

European Journal of Medicinal Plants 13(4): 1-14, 2016, Article no.EJMP.23930 ISSN: 2231-0894, NLM ID: 101583475



SCIENCEDOMAIN international www.sciencedomain.org

Antidiabetic Effects of *Pterocarpus marsupium* (*Gammalu*)

H. K. I. Perera^{1'}

¹Department of Biochemistry, Faculty of Medicine, University of Peradeniya, Sri Lanka.

Author's contribution

The sole author designed, analyzed, interpreted and prepared the manuscript.

Article Information

DOI: 10.9734/EJMP/2016/23930 <u>Editor(s)</u>: (1) Elena Maria Varoni, Dipartimento di Scienze Biomediche, Chirurgiche ed Odontoiatriche, University of Milan, Italy. (2) Marcello Iriti, Professor of Plant Biology and Pathology, Department of Agricultural and Environmental Sciences, Milan State University, Italy. (1) Anthony Cemaluk C. Egbuonu, Michael Okpara University of Agriculture Umudike, Nigeria. (2) Juei-Tang Cheng, Chang Jung Christian University, Taiwan. Complete Peer review History: <u>http://sciencedomain.org/review-history/13976</u>

Review Article

Received 29th December 2015 Accepted 11th March 2016 Published 1st April 2016

ABSTRACT

Herbal medicines are considered to offer gentle means of managing chronic diseases at a lower cost. Pterocarpus marsupium Roxb. (PM) (Gammalu in Sinhala) heartwood and bark have been used as antidiabetic remedies in many cultures for thousands of years. The aim of this review is to address the existing evidence on antidiabetic effects of the P. marsupium. The hypoglycaemic effects, antidyslipidaemic effects, antioxidative effects and the safety of the PM heartwood and the bark have been scientifically validated using a multitude of *in vitro* and *in vivo* studies. Multiple mechanisms responsible for hypoglycaemic effects of PM including β -cell regeneration, insulin release and insulin-like actions of some compounds isolated were identified. (-)-Epicatechin, a flavonoid isolated from the bark has shown insulin-like effects, effects on β -cell regeneration and insulin release. Several compounds including pterostilbene and marsupsin isolated from the PM heartwood were identified as compounds with hypoglycaemic effects. The latex (gum) of the tree is a popular remedy used in Sri Lanka for diabetes even though the literature on PM does not discuss about the antidiabetic effects of the latex. Few investigations focused on the antidiabetic effects of PM latex have demonstrated strong inhibitory effects of the latex on α -amylase and α -glucosidase activities and on protein glycation. Investigations focusing on the antidiabetic effects and possible toxicity of the PM latex are essential to validate its efficacy and safety.

Keywords: Pterocarpus marsupium; Gammalu; antidiabetic; latex.

1. INTRODUCTION

Humankind has a long history of using herbal remedies to treat diseases. Herbal medicines are considered to offer more gentle means of managing chronic diseases at a lower cost. Studies of traditional antidiabetic remedies around the world have identified more than 1,200 species of plants with hypoglycemic activity [1]. Due to their perceived effectiveness, fewer side effects in clinical experience and relatively low costs, herbal drugs are now being prescribed widely by many health practitioners [2]. Diabetes is characterized by hyperglycemia and absolute or relative deficiency of insulin action leading to the development of chronic diabetic complications [3]. These complications include retinopathy, nephropathy, neuropathy and cardiovascular diseases. The increase in the oxidative stress, dyslipidaemia and glycation of macromolecules are major mechanisms contributing to the development of diabetic complications. Therefore therapeutic agents which can normalize the metabolic alterations that occur in diabetes are invaluable in the treatment of diabetes. The aim of this review is to address the existing evidence on antidiabetic effects of the P. marsupium.

Published data were searched within the year range of 1950-2015. Main search words used were *Pterocarpus marsupium*, antidiabetic, diabetes, hypoglycaemic, antidyslipidaemic, antioxidant, antiglycation and toxicity. Key search engines employed were Google scholar and PubMed.

Pterocarpus marsupium Roxb. (PM) (Gammalu in Sinhala) is known as Indian kino and belongs to the family Fabaceae (Leguminoceae). PM is native to Sri Lanka, India and Nepal. It has a long history as a versatile medicinal plant with multiple pharmacological activities. PM is extensively used to treat diabetes mellitus for thousands of years [4-8]. Even the ancient renowned text, Charaka Samhita which is believed to be one of the oldest of the surviving ancient treatise of ayurveda recommended PM for treating diabetes mellitus [7,9]. PM grows as a medium to large tree with a height up to 30 meters. The outer bark of PM is rough and vertically cracked [9]. The inner heartwood is golden yellow in colour while the outer sapwood is light yellow. Leaves are compound and flowers are yellow. Fruits have a flat circular wing. Latex (gum), which looks like dried blood comes out through the bark when an incision is made up to the cambium [9].

Water kept overnight in the tumblers made out of the PM heartwood is a popular traditional remedy used to lower blood glucose concentration in diabetics [10-12]. Several commercial preparations which include PM as a therapeutic agent alone or in combinations are available to treat diabetes [13]. Even though the scientific evidence for the use of the PM latex as an antidiabetic agent is lacking, the latex obtained from PM is widely used to treat diabetes in Sri Lanka [14]. A dose of one tea spoon of gum daily is considered as the effective dose [15,16]. A patent was obtained for a Sri Lankan avurvedic preparation made with five antidiabetic plants to treat diabetes in which the main ingredient is PM latex [17]. Latex contains 75% tannic acid, kino tannic acid (a non-glucosidal tannin), kinonin, kinored and small quantities of catechol, protocatechic acid, resin pectin and gallic acid [9].

Hypoglycaemic effects of the PM heartwood and bark are studied extensively. Many of these studies were conducted in diabetes induced rats and there are some reports on the clinical trials conducted with PM. Some of these studies have addressed the mechanism of action (Fig. 1) and toxicity of the extracts as well. Evidence for the β -cell regeneration in the pancreas [18,19], insulin release [20,21], insulin-like action [20-24], increased expression of glucose transporter [22], inhibition of digestive enzymes amylase and glucosidase (by the latex) [25,26] are some of the mechanisms identified. Furthermore, evidence on protective effects such as antioxidant effects [7,27], antidyslipidaemic effects [27-29] and evidence for antiglycation effects of the latex are available [30,31]. Several compounds responsible for the antidiabetic effects of PM have been revealed. Many investigations have focused on pterostilbene and (-)-epicatechin which were recognized as two major compounds responsible for the antidiabetic effects of heartwood and bark respectively.

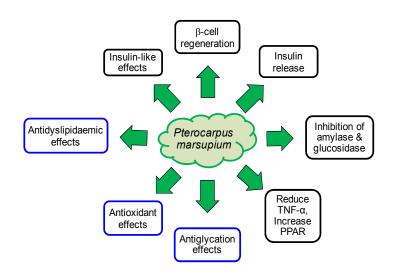


Fig. 1. Scientifically proven antidiabetic effects of Pterocarpus marsupium Effects outlined in black are responsible for hypoglycaemic effects. Additional effects outlined in blue provide further protection in alleviating diabetic complications

2. EXPERIMENTAL EVIDENCE ON HYPOGLYCAEMIC EFFECTS

Experiments conducted *in vivo* in animal models and humans have validated the hypoglycaemic effects of PM heartwood and bark as well as various fractions and compounds isolated from them.

2.1 Hypoglycaemic Effect of PM Heartwood

Methanol, ethanol and aqueous extracts of PM heartwood and their sub fractions have shown hypoglycaemic effects in streptozotocin or alloxan- induced rats. More recent studies have used a daily dose of 100 to 300 mg/kg body weight for 7 to 28 days which were effective. However, others have used higher doses such 750 mg or 1 g/kg for 5 to 30 days. Several clinical trials have demonstrated the hypoglycaemic effects of PM heartwood with a flexible dose of 2 to 4 g/day for a period of 12 or 36 weeks.

Different extracts prepared from the heartwood of PM (crude powder, ethanolic extract and sub fractions of ethanol extract; aqueous, chloroform, hexane and n-butanol fractions) resulted an improvement in the oral glucose tolerance after a sucrose load in normal rats [21]. This study revealed that, after a sucrose load, all fractions other than the aqueous fraction improved the oral glucose tolerance in streptozotocin-induced diabetic rats as well. Ethanol extract (100 mg/kg/day) was found to be effective when given to diabetic rats for 10 consecutive days [21]. Similar effect was observed with ethanol extracts of PM heartwood (150 mg/kg) administered daily for a period of 14 days in alloxan-induced diabetic rats. It resulted in a reduction in blood glucose [27]. Aqueous extract of the PM heartwood (100 and 200 mg/ kg/day) given orally for 4 weeks decreased the fasting and postprandial blood glucose concentration in streptozotozin-induced diabetic rats [32]. Methanol extract of PM heartwood (300 mg/kg/day) given for 7 and 14 days caused a reduction in serum glucose in streptozotocininduced diabetic rats [33]. When PM heartwood ethanol extract was given at 100, 200, 300 mg/kg/day for 12 days, it had shown a reduction in fasting blood glucose levels in streptozotocininduced diabetic rats [34]. Aqueous extract of PM heartwood (250 mg/kg/day) has produced hypoglycemic effects in alloxan-induced diabetic rats [35]. Methanol extract of PM heartwood (750 mg/kg/day for 6 days) caused a reduction in blood glucose concentration in both normal and diabetic rats [7]. Aqueous extract of PM bark (1 g/kg/day) resulted in a decrease in blood glucose by 38% and 60% on 15th and 30th day in streptozotozin-induced diabetic rats [1]. Ethyl acetate fraction of ethanol extract of PM heartwood lowered the blood sugar concentration in alloxan-diabetic rats treated for 5 days [36].

Pterostilbene isolated from PM heartwood (10 mg/kg/ intravenous) has caused hypoglycemia in dogs [37]. Marsupsin and pterostilbene have lowered the blood glucose level in a manner comparable to that of metformin when marsupsin, pterosupin and pterostilbene from the PM heartwood were given to streptozotocin-induced diabetic rats [38].

Antidiabetic effects of PM heartwood were explicitly demonstrated in clinical trials conducted in India [12]. In one such study, PM lowered postprandial blood fasting and glucose concentrations in 69% of the patients by the 12th wk [39]. In this group a dose of 2 g/ day was sufficient to produce glycaemic control in 73% of the patients while rest of the patients received either 3 or 4 g/day [39]. A similar double-blind trial conducted using capsulated dried decoction of PM heartwood (2-4 g/day) showed glycaemic control in 86% of patients treated with PM for 36 weeks [4]. In another clinical study PM was found to be useful in treating non-obese diabetic patients [40].

2.2 Hypoglycaemic Effect of PM Bark

Ethanol and aqueous extracts of PM bark were investigated in several studies. In one study, ethanol extracts of PM bark (150 mg/kg) administered daily for a period of 14 days to alloxan-induced diabetic rats caused a reduction in blood glucose [27]. Another study revealed that various sub fractions of the alcohol extract of PM bark were effective as antidiabetic agents in alloxan-induced diabetic rats and the butanol subfraction was found to be most effective [29]. Aqueous extracts of PM bark (1 g/kg/day) given to rats fed with a fructose-rich diet for 30 days lowered the serum glucose concentration compared to the group which did not receive the extract [28]. Aqueous extract of PM bark lowered blood glucose and improved glucose tolerance with no side effects in alloxan-induced diabetic rats [41]. In another study conducted with alloxan-induced diabetic rats, hypoglycaemic effects of a 21 day dose of aqueous extract of the PM stem bark (1 g/kg) was demonstrated [42].

A compound responsible for the hypoglycaemic effects was identified from the PM stem bark. First it was recognized that a flavonoid fraction from the PM bark effectively reduces the blood glucose concentration in alloxan induced-diabetic rats [18]. Another study conducted by the same group determined the hypoglycaemic effects of

the flavonoid (-)-epicatechin in alloxan-induced diabetic rats after administering 30 mg/kg/day for 4 to 5 days [19].

3. MECHANISM OF ACTION

3.1 β-cell Regeneration

There is evidence to suggest that (-)-epicatechin is effective in β -cell regeneration. Chakravarthy et al. [18] demonstrated the effectiveness of a flavonoid fraction from the PM bark in reversing the β -cell population of the pancreas in alloxaninduced diabetic rats. Later they showed that (-)epicatechin given for 4 to 5 days resulted in regeneration of the β -cell population of the islets of pancreas which were previously necrosed due to alloxan in alloxan-induced diabetic rats [19]. Functional nature of the regenerated β -cells was identified based on the immune reactive insulin studies [19]. However, some other investigators could not confirm the effectiveness of epicatechin in β -cell regeneration [43].

3.2 Insulin Release

Studies have revealed that ethanol and aqueous extracts of PM heartwood, their sub fractions and (-)-epicatechin from PM bark increase insulin release. Ethanol extract of PM heartwood given for 10 days increased serum insulin concentration in streptozotocin-induced diabetic rats [21]. Antidiabetic constituents in the aqueous extract of PM heartwood were fractionated using bioassay-guided fractionation in a study and a high molecular weight fraction (>10 < 30 kDa) isolated has shown potent insulinotrophic and insulin-like properties [20]. The same study revealed that PM stimulated the insulin secretion from the mouse pancreas in a concentrationdependent manner in vitro [20]. Furthermore, the increase in glucose clearance observed after intra venous administration of glucose to normoglycaemic sheep was positively correlated with insulin secretion when water extract and the high molecular weight fraction of PM were administered [20]. In another investigation, ethyl acetate fraction of the ethanol extract of PM heartwood increased blood insulin concentration in alloxan-diabetic rats treated for 5 days [36]. The same study showed that (-)-epicatechin in the water extract of PM bark increased the cAMP content of the islets of pancreas in rats with an associated increase in insulin release and conversion of proinsulin to insulin [36]. Ethanol extracts of PM heartwood and bark administered daily for a period of 14 days increased

the plasma insulin concentration in diabetic rats [27].

3.3 Insulin-like Activities

3.3.1 Increase in glucose uptake

Increase in glucose uptake by peripheral tissues is a major event which lowers the blood glucose concentration during fed state. Skeletal muscle and adipose tissue are major tissues which respond to insulin mediated increase in glucose uptake. Mishra et al. [21] studied the effects of five phenolic compounds isolated from n-butanol fraction of the ethanolic extract of PM heartwood on 2-deoxy-glucose uptake by mouse skeletal muscle cells (C2C12). When incubated for 24 h of which the final 3 h prior of the assay were in serum free media, four phenolic compounds namely vijayoside, pteroside, marsuposide and pterosupol (10 µM/ml) increased the glucose uptake in basal and insulin-stimulated cells in a concentration-dependent manner [21]. The other compound 2,4, dihydroxy-phenyl phenolic glucopyranosyl-C-glycoside did not show an effect on glucose uptake [21]. In another study, a high molecular weight fraction of PM heartwood aqueous extract stimulated glucose uptake by mouse skeletal muscle in a dose-dependent manner in vitro [20]. They also suggested that PM and insulin share a common signaling pathway, as the PM mediated muscle glucose uptake did not improve in the presence of insulin [20]. PM methanolic extract and an isoflavone purified from the extract have increased glucose uptake in L6 myotubes with a parallel increase in the GLUT4 expression which was comparable with those of insulin and rosiglitazone [22]. In this study, the fraction showing maximum glucose uptake was purified and the active compound was identified as 7-O-α-L-rhamnopyranosyl-oxy-4'-methoxy-5-hydroxy isoflavone [22]. Evidence from the same study suggested the involvement of a PI3 kinase dependent pathway by PM methanol extract and a PI3 kinase independent pathway by 7-O- α -L-rhamnopyranosyl-oxy-4'methoxy-5-hydroxy isoflavone in increasing glucose transport [22].

Evidence for the effect of (-)-epicatechin in increasing the glucose uptake in rat diaphragm in a dose-dependent manner is available [44].

3.3.2 Increase in glycogen synthesis

During the fed state excess glucose is channeled to synthesize glycogen with the action of insulin, resulting in lowering of blood glucose. Several investigations have looked in to the ability of PM in restoring glycogen synthesis. Aqueous extract of PM (1 g/kg/day) corrected the decline in glycogen content occurred in insulin dependent tissues such as liver and skeletal muscle of diabetic rats [1]. Oral administration of the aqueous extract of PM bark (300 and 500 mg/kg/day for 12 weeks) to diabetic rats restored the level of glycogen synthase [23]. (-)-Epicatechin increased the glycogen content in rat diaphragm in a dose-dependent manner [44]. Ahmad et al. revealed insulin-like activities of demonstrated (-)-epicatechin and that (-)-epicatechin does not share the plasma membrane receptor with insulin [44].

3.3.3 Increase in activity of oxidative enzymes

Glycolysis and tricarboxylic acid (TCA) cycle are central pathways which oxidize glucose and are important in the regulation of glucose homeostasis. Dysregulation of glycolysis occurs in diabetes due to reduction in the amount and/or activity of regulatory enzymes of glycolysis [45,46]. Reduction of TCA cycle flux in tissues such as skeletal muscle [24] and peripheral neurons [46] have been observed in diabetes. In diabetic rats the activities of glucokinase, hexokinase and phosphofructokinase were found to be reduced by 35%, 50% and 60% of the controls respectively [1]. Aqueous extract of PM g/kg/day) has corrected this alteration (1 completely in phosphofructokinase and partly in glucokinase and hexokinase of diabetic rats [1]. Oral administration of the aqueous extract of PM bark (300 and 500 mg/kg/day) to diabetic rats restored the levels of glucokinase, lactate dehydrogenase, succinate dehydrogenase and malate dehydrogenase at the end of 12th week [23]. Authors have suggested that the restoration of the enzyme levels after administration of PM might be due to stimulatory effects of the extract on insulin secretion [23]. In diabetic-induced rats, pterostilbene (40 mg/kg for 6 weeks) increased the activity of hexokinase. Furthermore, it decreased the activities of gluconeogenic enzymes glucose-6-phosphatase and fructose-1,6-bisphosphatase in the liver leading to a decrease in the release of glucose into the circulation [47].

3.4 α-amylase & α-glucosidase Inhibitory Effects

Antidiabetic drugs target at various biochemical mechanisms including digestion of

carbohydrates. α -Amylase and α -glucosidase are principle enzymes involved in digesting dietary carbohydrates into absorbable molecules. Inhibition of these enzymes is helpful in bringing down postprandial blood glucose spikes. Strong *in vitro* inhibitory effects of PM latex were observed on α -amylase and α -glucosidase with IC₅₀ of 2.97 and 0.54 µg/ml respectively [26]. Aqueous extract of the PM latex had shown a marked α -glucosidase inhibitory activity [25].

3.5 Effects on TNF-α and Peroxisome Proliferator Activator Receptors (PPARs)

The inflammatory cytokine tumor necrosis factor TNF- α is elevated in type 2 diabetes and it is known to cause insulin resistance [48]. TNF-α has a direct inhibitory effect on insulin signaling [49] in addition to indirect effects leading to insulin resistance. Aqueous extract of PM heartwood (100 and 200 mg/kg/day) given orally for 4 weeks decreased TNF- α to the normal levels in diabetic rats [32]. TNF-α down regulates peroxisome proliferator activator receptors (PPARs) which are nuclear receptors that regulate metabolic events of cells [50]. Methanolic extract of PM and an isolated 7-O-α-L-rhamnopyranosyl-oxy-4'isoflavone methoxy-5-hydroxy isoflavone induced the expression of PPAR-γ in L6 myotubes [22]. In the same study cycloheximide mediated inhibition of protein synthesis blocked the effect of the extract and the isoflavone suggested that new protein synthesis is required for these effects [22]. Furthermore, PI3 kinase was activated by the methanolic extract but not by the pure isoflavone suggesting the need of a PI3 kinase independent pathway for the effect of isoflavone [22]. It has been revealed that pterostilbene (100 µM) activates PPAR-a activity by 8 fold [51].

4. OTHER PROTECTIVE EFFECTS

4.1 Antidyslipidemic Effects

Diabetes leads to derangement in the lipid metabolism [52] which becomes an underlying cause for the increase in the risk of cardiovascular diseases in diabetes. Scientific evidence for antidyslipidaemic effects of PM is available. The ethanolic extract of PM (100 mg/kg) had demonstrated antidyslipidemic effects in hamsters that were on a high fat diet [21]. Lipid parameters (total cholesterol, LDL cholesterol, VLDL cholesterol, HDL cholesterol and triglycerides) were restored when ethanol extracts of PM heartwood and bark (150 mg/kg) were given to alloxan-induced diabetic rats for 14 days [27]. In another study, PM controlled the diabetes related metabolic alterations of cholesterol and triglycerides in diabetic rats [29]. Only extract had prevented PΜ the hypertriglyceridaemia when an aqueous extracts of PM bark (1 g/kg/day), Ocimum sanctum leaves and Trigonella foenum-graecum seeds were given to rats concurrently with a fructoserich diet for 30 days [28]. However, in this study, all three extracts have lowered blood glucose concentrations [28]. Hypoglycaemic effects of O. sanctum leaves were revealed in a recent study as well [53]. Ethyl acetate extract of PM heartwood and its flavonoid constituents, marsupsin, pterosupin and liquiritigenin were orally administered to hyperlipidaemia-induced rats for 14 days [54]. Ethyl acetate extract was found to be effective in reducing serum triglyceride, total cholesterol, LDL cholesterol and VLDL cholesterol concentration without changing HDI cholesterol [54]. Liquiritigenin and pterosupin reduced the serum cholesterol, LDL cholesterol and atherogenic index while pterosupin lowered serum triglyceride too [54]. Pterostilbene ppm) (25 given to hypercholesterolaemic hamsters reduced LDL cholesterol and increased HDL cholesterol [51].

4.2 Antioxidant Effects

Persistent hyperglycemia induces the oxidative stress in diabetic patients as a result of increased free radical generation and impaired antioxidant defense system [52,55]. Increased oxidative stress contributes to diabetic complications [56]. The antioxidant defense system includes the enzymes superoxide dismutase, catalase, glutathione peroxidase and glutathione reductase [57]. Reduced glutathione is a key endogenous non enzymatic antioxidant [57]. PM heartwood, bark and (-)-epicatechin from the bark have demonstrated protection against oxidative stress in diabetes.

Methanol extract of PM heartwood (750 mg/kg/day) administered for 6 days reversed the level of superoxide dismutase and catalase in streptozotocin-induced diabetic rats [7]. Antioxidant status was normalized when alloxaninduced diabetic rats were treated with ethanol extracts of PM heartwood and bark (150 mg/kg/day) for 14 days [27]. The same study demonstrated an improvement in the concentration of superoxide dismutase, catalase,

glutathione peroxidase, glutathione reductase and reduced glutathione [27].

Methanolic extract of PM bark showed antioxidant effects on H₂O₂ induced oxidative stress in isolated frog heart muscle as demonstrated by the time taken for cardiac arrest to occur [58]. Aqueous extract of PM stem bark showed a strong antioxidant activity in vitro as shown by measurements of DPPH scavenging and ferric reducing ability [59]. The study also demonstrated the normalization of the antioxidant status (superoxide dismutase, catalase, glutathione peroxidase and glutathione reductase) in the liver slices [59].

Level of reduced glutathione was found to be decreased in type 2 diabetes [6]. Insulin caused an increase in the reduced glutathione level of normal and type-2 diabetic erythrocytes. (-)-Epicatechin is a member of catechins a group of compounds widely known for their antioxidant effect. A remarkable increase in the glutathione content of normal and type-2 diabetic erythrocytes was observed with 1 mM and 10 mM (-)- epicatechin [6]. In diabetic-induced rats, pterostilbene (40 mg/kg for 6 weeks) increased the activity of superoxide dismutase, catalase, glutathione peroxidase and reduced glutathione [60].

4.3 Glycation Inhibitory Effects

Advanced glycation end products (AGEs) are produced as a result of non enzymatic glycation of molecules. AGEs are key players of causation of chronic diabetic complications. Persistent elevation of blood glucose existing in diabetes leads to formation of AGEs at an enhanced rate. A reduction of the level of early glycation product, glycosylated hemoglobin (HbA1c) was revealed with the administration of PM. However, it is not clear whether the decline seen was a direct effect or an indirect effect of the hypoglycaemic effects of PM. Ethanol extracts of PM heartwood and bark (150 mg/kg/day for 14 days) has significantly reduced HbA1c in alloxan-induced diabetic rats [27]. Methanol extract of PM heartwood (300 mg/ kg/ day for 7 and 14 days) lowered the HbA1c concentration by 14.4 and 22.6% respectively in diabetic rats [33]. PM (2-4 g/day) produced a 4% decline in HbA1c at 12 wks in type 2 diabetic patients [39]. Two recent in vitro studies conducted have demonstrated strong alvcation inhibitory effects [30] and glycation induced cross-link inhibitory effects [31] of PM latex in the presence of high concentration

of fructose. These in vitro studies found evidence for the possible beneficial effects of PM latex in reducing diabetic complications [30,31]. (-)-Epicatechin was able to break preformed glycated human serum albumin *in vitro* [61]. Furthermore (-)-epicatechin (50 and 100 mg/kg for two weeks) reduced AGE accumulation in the retinas of rats injected with AGEs [61].

4.4 Anti-cataract Effects

Diabetes increases the risk of developing cataract. An aqueous extract of PM bark (1 g/kg), aqueous extract of *Ocimum sanctum* leaves and alcoholic extract of *Trigonella foenum-graecum* seeds were tested in alloxan-induced diabetic rats that have developed cataract. The best anti-cataract effects were observed with PM in this study [62].

5. COMPOUNDS WITH ANTIDIABETIC PROPERTIES

Several polyphenolics isolated from PM have showed antidiabetic effects. Pterostilbene, 7-O-α-L-rhamnopyranosyl-oxy-4'marsupsin, methoxy-5-hydroxy isoflavone, marsuposide, pteroside, pterosupol, vijayoside, liquiritigenin and pterosupin isolated from PM heartwood and (-)-epicatechin isolated from PM bark are recognized as antidiabetic compounds (Table 1, Fig. 2). Concentration of pterostilbene was demonstrated to be several times higher than that of marsupsin [63]. A recovery of 17.5% epicatechin is evident in 50% methanol water extract of PM bark [64]. Furthermore, there is evidence for the presence of a high molecular weight antidiabetic compound (>10<30 kDa) in PM heartwood [20]. Accordingly, pterostilbene from heartwood and (-)-epicatechin from bark seem to be two major antidiabetic compounds of PM.

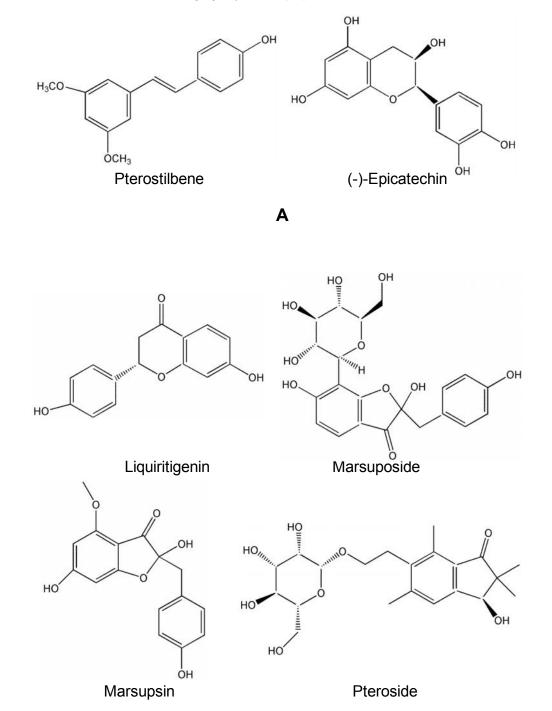
6. TOXICITY

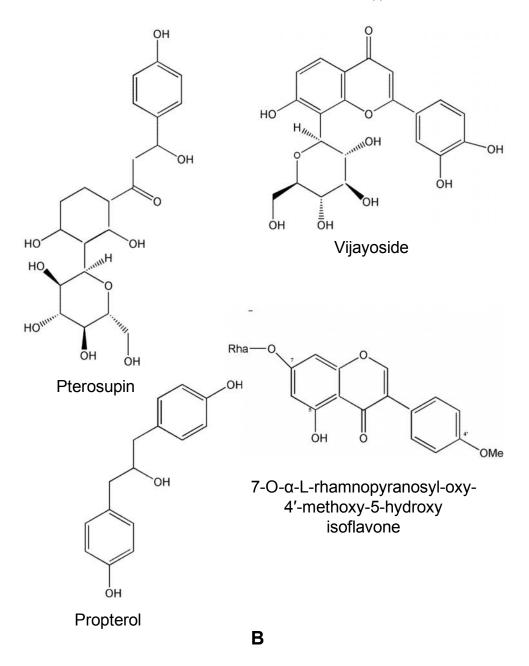
Commonly used herbal medicines which have been in use for thousands of years are less likely to cause significant toxic effects. There is scientific evidence to support the safety of PM heartwood, bark and pterostlibene demonstrating lack of hepatic and renal toxicity. Some studies have shown hepatoprotective effects of PM as well.

Evidence from a clinical trial revealed the lack of toxic effects in type 2 diabetic patients when a

dose of 2-4 g PM per day was given for 36 weeks [4]. The altered renal and hepatic function markers were normalized when ethanolic extract of PM heartwood (100 mg/kg) was given for 28 consecutive days to diabetic rats that were on a high fat diet [21]. Ethanol extract of PM heartwood (5, 50, 100 and 1000 mg/kg/day) did

not show acute toxicity in alloxan-induced diabetic rats when administered using gastric intubations and observed for another 14 days [27]. Normalization of alanine transaminase, aspartate transaminase and alkaline phosphatase levels was observed in this study [27]. Methanol extract of PM heartwood did not







A: Two major compounds of PM with antidiabetic effects, B: Other antidiabetic compounds identified from PM. (Identity of propterol with pterosupol was revealed previously [65]. Structures were adapted from published data [66,67] except for 7-O-α-L-rhamnopyranosyl-oxy-4'-methoxy-5-hydroxy isoflavone)

show acute toxicity in mice after oral administration doses of 500, 1000, 2000, 4000 and 8000 mg/kg and no mortality was observed up to 7 day in mice [7]. Methanol and water extracts of heartwood had reversed the hepatotoxic effects of CCl_4 as shown by the

normalization of serum bilirubin, transaminases and normal histology in liver [68]. Pterostlibene when given at a dose of 125 mg twice daily for 6-8 weeks to hypercholesterolaemic patients did not show signs of hepatic or renal toxicity [69].

Source	Compound	Effect- mechanism
PM heartwood	Pterostilbene	PM Heartwood Hypoglycaemic [37,38]- Increase
		glucose oxidation [47], decrease gluconeogenesis
		[47], peroxisome proliferator activator receptor
		agonist [51], antidyslipidaemic [51], antioxidant [59]
	7-O-α-L-rhamnopyranosyl-	Hypoglycaemic- Increase glucose uptake [22],
	oxy-4'-methoxy-5-hydroxy	GLUT4 expression [22], peroxisome proliferator
	isoflavone	activator receptor expression [22]
	Marsuposide	Hypoglycaemic- Increase glucose uptake [21]
	Marsupsin	Hypoglycaemic [38]
	Pteroside	Hypoglycaemic- Increase glucose uptake [21]
	Pterosupol (Propterol) [65]	Hypoglycaemic- Increase glucose uptake [21]
	Vijayoside	Hypoglycaemic- Increase glucose uptake [21]
	Liquiritigenin	Antidyslipidaemic [54]
	Pterosupin	Antidyslipidaemic [54]
PM bark	(-)-Epicatechin	Hypoglycaemic [19]- β-cell regeneration [19],
		increase in insulin release [36], conversion of
		proinsulin to insulin [36], glucose uptake [44] and
		glycogen synthesis [44]
		Protective effects- antioxidant [6], antiglycation [60]

Table 1. Antidiabetic compounds of PM heartwood and bark and their mechanisms

PM bark was also able to control the diabetes related increases in transaminases and alkaline phosphatase levels [29]. Methanol and aqueous extracts of PM stem bark (25 mg/kg/day for 14 days) showed a significant hepatoprotective activity against CCl₄ induced hepatotoxicity in rats with a more pronounced effect with the methanol extract [70]. The hepatic damage seen in streptozotocin-induced diabetic rats normalized with the treatment of PM stem bark methanol extract at a dose of 100 and for 21 days, 300 mg/Kg/day indicating hepatoprotective effects of PM [71].

7. CONCLUSION

Pterocarpus marsupium (Gammalu in Sinhala) heartwood and bark have been extensively used in the treatment of diabetes in many cultures for thousands of years. This review has focused on the scientific evidence available on antidiabetic effects of P. marsupium and is presented highlighting the existing evidence for antidiabetic effects under different cellular mechanisms. Existing evidence validates the use of P. marsupium to treat diabetes and demonstrated the hypoglycaemic effects. antidyslipidaemic effects, antioxidative effects and the safety of the P. marsupium. This review revealed that the investigations were focusing on the heartwood, bark and some compounds present in these two parts. It was identified that the scientific evidence on the antidiabetic effects of the latex of P. marsupium is meager even though it is a popular remedy used in Sri Lanka for diabetes. Hence it is necessary to scientifically validate the efficacy and the safety of the *P. marsupium* latex.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

ACKNOWLEDGEMENTS

Prof. R. Sivakanesan and Mrs. Chintha Ellawala, Coordinator/ English Language Teaching Unit for their valuable comments in revising the manuscript. Prof. Lalith Jayasinghe for preparing the structure of $7-O-\alpha$ -l-rhamnopyranosyloxy-4'-methoxy-5-hydroxy isoflavone and Mr. A.M.P.S.T.M Bandara for drawing other chemical structures based on published figures.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. Grover JK, Vats V, Yadav S. Effect of feeding aqueous extract of

Pterocarpus marsupium on glycogen content of tissues and the key enzymes of carbohydrate metabolism. Molecular and Cellular Biochemistry. 2002;241(1-2): 53-59.

- Haque N, Salma U, Nurunnabi TR, Uddin MJ, Jahangir MFK, Islam SMZ, et al. Management of type 2 diabetes mellitus by lifestyle, diet and medicinal plants. Pakistan Journal of Biological Sciences. 2011;14:13-24.
- Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circulation Research. 2010;107(9):1058-1070.
- Hariharan RS, Venkataraman S, Sunitha P, Rajalakshmi S, Samal KC, Routray BM. Efficacy of vijayasar (*Pterocarpus marsupium*) in the treatment of newly diagnosed patients with type 2 diabetes mellitus: A flexible dose double-blind multicenter randomized controlled trial. Diabetologia Croatica. 2005;34(1):13-20.
- 5. Jung M, Park M, Lee HC, Kang YH, Kang ES, Kim SK. Antidiabetic agents from medicinal plants. Current Medicinal Chemistry. 2006;13(10):1203-1218.
- Rizvi SI, Zaid MA. Intracellular reduced glutathione content in normal and type 2 diabetic erythrocytes: Effect of insulin and (-) epicatechin. Journal of Physiology and Pharmacology. 2001;52(3):483-488.
- Joshi MC, Dorababu M, Prabha T, Kumar MM, Goel RK. Effects of *Pterocarpus marsupium* on NIDDM-induced rat gastric ulceration and mucosal offensive and defensive factors. Indian Journal of Pharmacology. 2004;36(5):296.
- Warrier PK, Nambiar VPK, Ramankutty C (eds). Indian medicinal plants. A compendium of 500 species. Chennai: Orient Longman Ltd. 1995;381-383.
- Badkhane Y, Yadav AS, Sharma AK, Raghuwanshi DK, Uikey SK, Mir FA, et al. *Pterocarpus marsupium* Roxb-Biological activities and medicinal properties. International Journal of Advances in Pharmaceutical Sciences. 2010;1(4):350-357.
- Maheshwari JK, Singh KK, Saha S. Ethnomedicinal uses of plants by Tharus in Kheri Distt Uttar Pradesh. Bulletin of Medico-Ethno-Botanical Research. 1980;1: 318-337.
- 11. Evans WC. Trease and evans pharmacognosy, 15th ed. Elsevier Limited, Philadelphia. 2002;414-420.

- 12. Chaudhury RR. Antidiabetic effect of Vijayasar *Pterocarpus marsupium*. In Gupta SK, ed. Pharmacology and Therapeutics in the New Millennium, New Delhi: Narosa. 2001;355-356.
- 13. Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TPA. Indian herbs and herbal drugs used for the treatment of diabetes. Journal of Clinical Biochemistry and Nutrition. 2007;40(3):163-173.
- 14. Sri Bharathie KP. Non-wood forest products in Asia. Regional office for Asia and the Pacific publication. Food and Agriculture Organization of the United Nations, Bangkok. 1994;118.
- 15. Jayasekara DC. Arka prakaranaya, Colombo: Siriwardhana printers.1950;111.
- 16. Ediriweera ER, Ratnasooriya WD. A review on herbs used in treatment of diabetes mellitus by Sri Lankan ayurvedic and traditional physicians. Ayu. 2009; 30(4):373-391.
- 17. Hettigoda V. Ayurvedic composition for diabetes. 2001; WO2001072316 A1.
- Chakravarthy BK, Saroj G, Gambhir SS, Gode KD. Pancreatic beta cell regeneration–A novel antidiabetic mechanism of *Pterocarpus marsupium* roxb. Indian Journal of Pharmacology. 1980;12(2):123-127.
- Chakravarthy BK, Gupta S, Gode KD. Functional beta cell regeneration in the islet of pancreas in alloxan induced diabetic rats by (-)epicatechin. Life Sciences. 1982;31:2693-2697. DOI: 10.1016/0024-3205(82)90713-5
- Mohankumar SK, O'Shea T, McFarlane JR. Insulinotrophic and insulin-like effects of a high molecular weight aqueous extract of *Pterocarpus marsupium* Roxb. hardwood. Journal of Ethnopharmacology. 2012;141(1):72-79.
- 21. Mishra A, Srivastava R, Srivastava SP, Gautam S, Tamrakar AK, Maurya R, et al. Antidiabetic activity of heart wood of *Pterocarpus marsupium* Roxb. and analysis of phytoconstituents. Indian Journal of Experimental Biology. 2013; 51(5):363-374.
- Anandharajan R, Pathmanathan K, Shankernarayanan NP, Vishwakarma RA, Balakrishnan A. Upregulation of GLUT-4 and PPARγ by an isoflavone from *Pterocarpus marsupium* on L6 myotubes: A possible mechanism of action. Journal of Ethnopharmacology. 2005;97(2):253-260.

- Gayathri M, Kannabiran K. Studies on the ameliorative potential of aqueous extract of bark of *Pterocarpus marsupium* Roxb in streptozotocin- induced diabetic rats. Journal of Natural Remedies. 2010; 10(1):36-43.
- 24. Gaster M, Nehlin JO, Minet AD. Impaired TCA cycle flux in mitochondria in skeletal muscle from type 2 diabetic subjects: Marker or maker of the diabetic phenotype? Archives of Physiology and Biochemistry. 2012;118(3):156-189.
- 25. Abesundara KJ, Matsui T, Matsumoto K. Alpha-glucosidase inhibitory activity of some Sri Lanka plant extracts, one of which, *Cassia auriculata*, exerts a strong antihyperglycemic effect in rats comparable to the therapeutic drug acarbose. Journal of Agricultural and Food Chemistry. 2004;52(9):2541-2545.
- Poongunran J, Perera HKI, Fernando WIT, Jayasinghe L, Sivakanesan R. αglucosidase and α-amylase inhibitory activities of nine Sri Lankan antidiabetic plants. British Journal of Pharmaceutical Research. 2015;7(5):365-374.
- Maruthupandian VR. 27. Α, Mohan Antidiabetic, antihyperlipidaemic and antioxidant activity Pterocarpus of marsupium Roxb. in alloxan induced diabetic rats. International Journal of Pharm Tech Research. 2011;3(3):1681-1687.
- 28. Grover JK, Vats V, Yadav SS. *Pterocarpus marsupium extract (Vijayasar)* prevented the alteration in metabolic patterns induced in the normal rat by feeding an adequate diet containing fructose as sole carbohydrate. Diabetes, Obesity and Metabolism. 2005;7(4):414-420.
- 29. Dhanabal SP, Kokate CK, Ramanathan M, Kumar EP, Suresh B. Hypoglycaemic activity of *Pterocarpus marsupium* Roxb. Phytotherapy Research. 2006:20(1):4-8.
- 30. Perera HKI, Handuwalage CS. Detection of protein glycation inhibitory potential of nine antidiabetic plants using a novel method. Asian Journal of Medical Sciences. 2014;6(2):1-6.
- 31. Perera HKI, Handuwalage CS. Analysis of glycation induced protein cross-linking inhibitory effects of some antidiabetic plants and spices. BMC Complementary and Alternative Medicine. 2015;15:175. DOI: 10.1186/s12906-015-0689-1
- 32. Halagappa K, Girish HN, Srinivasan BP. The study of aqueous extract of

Pterocarpus marsupium Roxb. on cytokine TNF- α in type 2 diabetic rats. Indian Journal of Pharmacology. 2010;42(6):392.

- Gupta R, Gupta RS. Effect of *Pterocarpus* marsupium in streptozotocin-induced hyperglycemic state in rats: Comparison with glibenclamide. Diabetologia Croatica. 2009;38(2):39-45.
- Patil UK. Antidiabetic activity of the ethanolic extract of heartwood of bijasar (*Pterocarpus marsupium* roxb.) in streptozotocin-nicotinamide induced type 2 diabetic rats. African Journal of Traditional, Complementary and Alternative medicines; 2008.

DOI:http://dx.doi.org/10.4314/ajtcam.v6i0.7 85

- Mukhtar HM, Ansari SH, Ali M, Bhat ZA, Naved T. Effect of aqueous extract of *Pterocarpus marsupium* wood on alloxaninduced diabetic rats. Die Pharmazie-An International Journal of Pharmaceutical Sciences. 2005;60(6):478-479.
- Ahmad F, Khan MM, Rastogi AK, Kidwai JR. Effect of (-)epicatechin on cAMP content, insulin release and conversion of proinsulin to insulin in immature and mature rat islets. Indian Journal of Experimental Biology. 1991;29:516-520.
- Haranath PS, Ranganthrao K, Anjaneyulu CR, Ramnathan JD. Studies on the hypoglycemic and pharmacological actions of some stilbenes. Indian Journal of Medical Sciences. 1958;12:85-89.
- Manickam M, Ramanathan M, Jahromi MAF, Chansouria JPN, Ray AB. Antihyperglycemic activity of phenolics from *Pterocarpus marsupium*. Journal of Natural Products. 1997;60(6):609-610.
- Indian Council of Medical Research (ICMR) Collaborating Centres, New Delhi. Flexible dose open trial of Vijayasar in cases of newly diagnosed non-insulindependent diabetes mellitus. Indian Journal of Medical Research. 1998;108: 24-29.
- 40. Upadhyay OP, Singh RM Dutta K. Studies on antidiabetic medicinal plants used in Indian folk-lore. Aryavaidyan. 1996;9:159-167.
- 41. Pandey MC, Sharma VP. Hypoglycemic effect of bark of *Pterocarpous marsupium* Roxb. (Bijaka) on alloxan induced diabetes. Medicine and Surgery. 1975;15: 21.
- 42. Vats V, Grover J K, Rathi SS. Evaluation of anti-hyperglycemic and hypoglycemic

effect of *Trigonella foenum-graecum* Linn, *Ocimum sanctum* Linn and *Pterocarpus marsupium* Linn in normal and alloxanized diabetic rats. Journal of Ethnopharmacology. 2002;79(1):95-100.

- Sheehan EW, Stiff DD, Duah F, Slatkin DJ, Schiff PL, Zemaitis MA. The lack of effectiveness of (−)-epicatechin against alloxan induced diabetes in Wistar rats. Life Sciences. 1983;33(7):593-597.
- 44. Ahmad F, Khalid P, Khan MM, Rastogi AK, Kidwai JR. Insulin like activity in (-) epicatechin. Acta Diabetologica Latina. 1989;26:291-300.
- 45. Guo X, Li H, Xu H, Woo S, Dong H, Lu F, et al. Glycolysis in the control of blood glucose homeostasis. Acta Pharmaceutica Sinica B. 2012;2(4):358-367.
- 46. Hinder LM, Vivekanandan-Giri A, McLean LL, Pennathur S, Feldman EL. Decreased glycolytic and tricarboxylic acid cycle intermediates coincide with peripheral nervous system oxidative stress in a murine model of type 2 diabetes. Journal of Endocrinology. 2013;216(1):1-11.
- Pari L, Satheesh MA. Effect of pterostilbene on hepatic key enzymes of glucose metabolism in streptozotocin-and nicotinamide-induced diabetic rats. Life Sciences. 2006;79(7):641-645.
- Hu FB, Meigs JB, Li TY, Rifai N, Manson JE. Inflammatory markers and risk of developing type 2 diabetes in women. Diabetes. 2004;53(3):693-700.
- 49. Pickup JC. Inflammation and activated innate immunity in the pathogenesis of type 2 diabetes. Diabetes Care. 2004; 27(3):813-823.
- 50. Berger JP, Akiyama TE, Meinke PT. PPARs: Therapeutic targets for metabolic disease. Trends in Pharmacological Sciences. 2005;26(5):244-251.
- 51. Rimando AM, Nagmani R, Feller DR, Yokoyama W. Pterostilbene, a new agonist for the peroxisome proliferator-activated receptor α-isoform, lowers plasma lipoproteins and cholesterol in hypercholesterolemic hamsters. Journal of Agricultural and Food Chemistry. 2005; 53(9):3403-3407.
- Sinah PK. Banerjee S. 52. Baxi D, Ramachandran Therapy with AV. methanolic extract of Pterocarpus marsupium Roxb and Ocimum sanctum Linn reverses dyslipidemia and oxidative stress in alloxan induced type I diabetic rat

model. Experimental and Toxicologic Pathology. 2012;64(5):441-448.

- 53. Hannan JMA, Ojo OO, Ali L , Rokeya B, Khaleque J, Akhter M, et al. Actions underlying antidiabetic effects of *Ocimum sanctum* leaf extracts in animal models of type 1 and type 2 diabetes. European Journal of Medicinal Plants. 2015;5(1):1-12.
- 54. Jahromi MF, Ray AB, Chansouria JPN. Antihyperlipidemic effect of flavonoids from *Pterocarpus marsupium*. Journal of Natural Products. 1993;56(7):989-994.
- 55. Ahmed RG. The physiological and biochemical effects of diabetes on the balance between oxidative stress and antioxidant defense system. Medical Journal of Islamic World Academy of Sciences. 2005;15:31-42.
- 56. Maritim AC, Sanders RA,Watkins 3 JB. Diabetes, oxidative stress, and antioxidants: A review. Journal of Biochemical and Molecular Toxicology. 2003;17:24-38.
- 57. Chandra M, Chandra N, Agrawal R, Kumar A, Ghatak A, Pandey VC. The free radical system in ischemic heart disease. International Journal of Cardiology. 1994;43:121-125.
- 58. Rajyalakshmi G, Radhika T, Prasad N. Antioxidant activity of red Kino tree using frog heart model. Pharmacology Online. 2008;3:26-31.
- 59. Mohammadi M, Khole S, Devasagayam TPA, Ghaskadbi S. *Pterocarpus marsupium* extract reveals strong in vitro antioxidant activity. Drug Discovery and Therapeutics. 2009;3(4):151-161.
- 60. Satheesh MA, Pari L. The antioxidant role of pterostilbene in streptozotocinnicotinamide-induced type 2 diabetes mellitus in Wistar rats. Journal of Pharmacy and Pharmacology. 2006; 58(11):1483-1490.
- 61. Kim J, Kim CS, Moon MK, Kim JS. Epicatechin breaks preformed glycated serum albumin and reverses the retinal accumulation of advanced glycation end products. European Journal of Pharmacology. 2015;748:108-114.
- 62. Vats V, Yadav SP, Biswas NR, Grover JK. Anti-cataract activity of *Pterocarpus marsupium* bark and *Trigonella foenumgraecum* seeds extract in alloxan diabetic rats. Journal of Ethnopharmacology. 2004;93(2):289-294.

- 63. Mallavadhani UV, Sahu G. Pterostilbene: A highly reliable quality-control marker for the ayurvedic antidiabetic plant bijasar'. Chromatographia. 2003;58(5-6):307-312.
- 64. Kishore AR, Veerasamy R, Jain M. HPLC method for estimation of (-)-epicatechin in *Pterocarpus marsupium* herbal extracts and pharmaceutical dosage formulations. Eurasian Journal of Analytical Chemistry. 2010;6(1):31-39.
- Maurya R, Ray AB, Chattopadhyay SK, Duah FK, Lin MC, Slatkin DJ, Schiff Jr PL. The synthesis of propterol, a novel 1, 3-diarylpropan-2-ol from *Pterocarpus marsupium*. Journal of Natural Products. 1985;48(2):313-315.
- 66. Sigma catalogue. Available:<u>http://www.sigmaaldrich.com/catalog/product/sigma/p1499?lang=en®ion=LK</u> Available:<u>http://www.sigmaaldrich.com/catalog/product/aldrich/855235?lang=en®i</u>

on=LK 67. Pubchem. Available:<u>https://pubchem.ncbi.nlm.nih.gov</u> /search/search.cgi

- 68. Rane A, Grampurohit ND. Hepatoprotective activity of *Pterocarpus marsupium* and Butea koen-ex-Roxb. Indian Journal of Pharmaceutical Sciences. 1998;5:182-184.
- Riche DM, McEwen CL, Riche KD, Sherman JJ, Wofford MR, Deschamp D, Griswold M. Analysis of safety from a human clinical trial with pterostilbene. Journal of toxicology; 2013. Available:<u>http://dx.doi.org/10.1155/2013/46</u> 3595
- Mankani KL, Krishna V, Manjunatha BK, Vidya SM, Singh SDJ, Manohara YN, et al. Evaluation of hepatoprotective activity of stem bark of *Pterocarpus marsupium* Roxb. Indian Journal of Pharmacology. 2005;37(3):165-168.
- 71. Gupta R, Gupta RS. Hepatoprotective action of *Pterocarpus marsupium* against streptozotocin–induced oxidative stress. Egyptian Journal of Biology. 2010;12(1): 44-51.

© 2016 Perera; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://sciencedomain.org/review-history/13976