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# **Metabolism, Toxicity, Detoxification, Occurrence, Intake and Legislations of Fumonisins - A Review**

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#### *Authors' contributions*

*This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.*

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# **ABSTRACT**

Fumonisins are a group of mycotoxins generated by the *Fusarium spp*. in foods and feeds. More than 15 isomers of Fumonisin are recognized, and the B series of Fumonisins is the primary and referral isomer of Fumonisin. Fumonisin B can cause leukoencephalomalacia in rabbits and horses and porcine pulmonary edema in swine. Fumonisin B is also nephrotoxic, hepatotoxic, immunotoxic and carcinogenic. It blocks sphingolipid biosynthesis (and hinders the synthesis of ceramide) by a noticeable resemblance to sphingosine and sphinganine. This paper provides a review of the toxicity, occurrence, and mechanism of carcinogenicity, hepatotoxicity, nephrotoxicity as well as immunotoxicity of Fumonisins, which are primarily found on a variety of food and feed in Africa, America, Europe, Asia, and Oceania. In this paper, current information on contamination of feeds and foods by Fumonisins around the world is summarized. Because of economic losses induced by Fumonisins and their harmful effects on animal and human health, various procedures to detoxify infected feeds and foods have been illustrated in this review, including biological, physical, and chemical processes. Additionally, we discuss dietary intakes and maximum limits of Fumonisins in some countries.

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*Keywords: Fumonisins; toxicity; detoxification; mechanism; occurrence; intake.*

#### **1. INTRODUCTION**

Fumonisins are a group of more than ten mycotoxins created by *Fusarium* species like *F. globosum, F. oxysporum, F. proliferatum, F. verticillioides* and other species of *Fusarium, Alternaria alternata f. sp. lycopersici,* as well as *Aspergillus niger* [1,2].

Fumonisins have a noncyclic structure (in contrast to most mycotoxins), in which there is a chain with 19- or 20- carbon

aminopolyhydroxyalkyl, diesterified by tricarballylic acid groups (propane-1,2,3 tricarboxylic acid) as shown in Fig 1. Hitherto, various chemically associated series or groups of Fumonisins were isolated. These series consist of A, B, C, and P. The main detected forms of Fumonisins in foods, are the B series of Fumonisins [3]. In more than 15 Fumonisin forms, Fumonisins  $B_1$ , Fumonisins  $B_2$ , and Fumonisins  $B_3$  are the broadest mycotoxins that have been described [4].







**Fig. 1. Chemical structures of the Fumonisins [3,4]**

Fungi-producing Fumonisin contaminates the following: apple, barley, beef, breakfast cereals, black tea, corn, cornbread, corn flour, corn flakes, corn grits, corn snacks, basmati rice, crunchy nut, egg, milk, oats, polenta, popcorn, row corn, soybean, canned foods, tomato, tortilla, and wheat [5].

Intake of fumonisin B induced a different form of toxic effect on animals, including leukoencephalomalacia in horses [6], change in weight of the body and internal organ in broiler chicken [7,8], and pulmonary edema as well as hepatocellular necrosis in piglet [9,10]. Moreover, renal and hepatic toxicity are detected in different animals, such as rabbits, lambs, turkeys, mice, rats, and broilers [7,11-14].

In human, the risk of neural tube defects (NTD) and developing esophageal cancer is increased by Fumonisins [15,16].

# **2. METHODS**

The current review is a narrative on Fumonisins, and the databases including Science Direct, PubMed, and Google Scholar are used to collect the published articles since 1980 through 2018. This paper is conducted using keywords: [*Fumonisin* OR *mycotoxin* OR *Fumonisin B* OR *Fusarium*] AND [toxicity OR detoxification OR degradation OR mechanism OR metabolism OR occurrence OR prevalence OR intake OR limitation]. The list of references of included articles was also searched to identify additional ones. After first screening by the title and abstract, eligible studies were downloaded and easy and suitable phrases were chosen. Inclusion criteria in our study included: (1) Fulltext available. (2) Review, mini-review, original, narrative articles, and books. (3) Published paper in the English language (to avoid mistakes in translation process) among 1980 to the 2018 year. (4) Detect concentration of Fumonisin B1, B2, and B3 in barley, wheat, oat, rice, corn and corn product such as corn grits, corn flakes, corn flour, cornmeal, and corn kernel. The articles were excluded when they did not meet these criteria.

# **3. OCCURRENCE**

Due to increase in global grain exchange, the fungi spread was transferred from one country to another [17]. In *Fusarium* fungi, this hazard is expected to be minimum whereas these phytopathogens are field sooner than storage organisms. Table 1 describes the global infection of animal feeds and foodstuffs with Fumonisins is described.

#### **3.1 North and South America**

Corn is the most prevalent source of Fumonisins (Table 1). The level of Fumonisins in South America is higher than North America, maybe because of their different climate conditions. For instance, the concentration of Fumonisin in corn in Brazil reaches to 38.5 mg/kg [18], while the percentage of Fumonisins in corn product of North America such as corn flour, corn grits, and corn flakes rarely reaches to 1 mg/kg. This decline probably proves that detoxification method was more effective for the controlling Fumonisins in North America in comparison with South America.

In Brazil, the incidence of Fumonisins in corn was detected by [18-24], and the contamination of corn with Fumonisins in Brazil usually decreased over 1999 to 2014.

The average of Fumonisins in corn of Argentina was 10200 µg/kg in 2003 and 4700 µg/kg in 2004 [25].

The infection of wheat, oat and barely by Fumonisins was also detected by Mallmann et al. [23].

#### **3.2 Asia and Oceania**

In China, the contamination of corn with Fumonisins was reported by GAO and Yoshizawa., [36], Gong et al. [39], Li et al. [33], Li et al. [40], Shi et al. [41], Ueno et al. [37], and Yoshizawa. [35]. Based on these studies the most extreme concentration of Fumonisin  $B_1$ ,  $B_2$ and  $B_3$  were 25.97 mg/kg, 6.77 mg/kg and 4.13 mg/kg respectively. Also, Zhang et al. [38] reported that in China total Fumonisins concentration was 0.5 to 16 mg/kg. In Iran, Shephard et al., [44] investigated infection of corn with fumonisin  $B_1$ ,  $B_2$ , and  $B_3$ . Also, Alizadeh et al. [45] reported the corn's contamination with fumonisin  $B_1$ . The high concentration of Fumonisins in corn of Iran and China, justify the high prevalence of esophageal cancer in Iranian and Chinese people.

The contamination of corn with Fumonisin  $B_1$  and B<sub>2</sub> were detected by Ueno et al. [46] in Japan.

Bryden et al. [47] declared the measure of total Fumonisins in corn of Philippines and Vietnam was 0.3 to 10 mg/kg and 0.3 to 9.1 mg/kg, respectively.



# **Table 1. Occurrence of Fumonisins from human foods, cereals, and crops in various countries**







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<b>Nation-seed</b>	Fumonisin $B_1$ (mg/kg)	Fumonisin $B_2$ (mg/kg)	Fumonisin $B_3$ (mg/kg)	Reference
Kenya	0.11 to 12			[94]
<b>Corn meal</b>				
Argentina	0.06 to 2.86	0.061 to 1.09	0.018 to 1.015	$[72]$
Argentina	0.603 to 1.171	0.717		[89]
<b>Brazil</b>	0.56 to 4.93	0.21 to 1.38		$[73]$
Canada	0.05			[86]
Peru	0.66	0.13		[86]
<b>USA</b>	Average: 1	0.3		[86]
<b>USA</b>	Total Fumonisins: <0.25 to >1			$[74]$
China	$< 0.5$ to 8.8	$< 0.5$ to 2.8	$< 0.5$ to 0.9	[95]
Turkey	0.25 to 2.66	0.55		[85]
South Africa	Average: 0.14	Average: 0.08		[86]
Oat				
<b>Brazil</b>	0.17			$[23]$
UK	<b>Total Fumonisins not detected</b>			$[29]$
<b>Rice</b>				
Iran	21.59			$[45]$
<b>China 1999</b>	3.410 to 16.79			$[96]$
<b>China 2010</b>	0.0001 to 0.00164			[96]
<b>China 2014</b>	0 to $0.74$			[96]
UK	<b>Total Fumonisins not detected</b>			$[29]$
Wheat				
Brazil	24.35			$[23]$
Argentina_flour	0.0003	0.00124		[97]
France	Not detected			$[27]$
Spain	0.2 to 8.8	0.2		$[28]$
UK	<b>Total Fumonisins not detected</b>			$[29]$

Contamination of Taiwan's corn with Fumonisins was investigated by Rheeder et al. [49], Tseng and Liu. [32], and Yoshizawa et al. [48]. Increasing level of Fumonisins in Taiwan's corn declared that legislation and control programs in this country were not efficient and changing it seems necessary.

#### **3.3 Europe**

Doko et al. [51] published a review article on some information about the occurrence of Fumonisins from some European nations (Croatia, Poland, Portugal, and Romania). The highest concentration of Fumonisins in Croatia was 25.2 mg/kg, and the mean value was 4.509 mg/kg [52].

In Spain, contamination of corn with Fumonisins investigated by Arino et al. [57], Castellá et al. [55], and Sanchis et al. [54]. Also, Castella et al. [28] reported the concentration of Fumonisin  $B_1$ and  $B_2$  in wheat and barley. However, Fumonisin  $B_1$  was not found in wheat and barley of France [27].

Lew et al. [50] reported the corn contamination with fumonisin  $B_1$  in Austria.

In oat, barley and wheat of United Kingdom Patel et al. [29] have not detected Fumonisins but Preis and Vargas. [59] declared the concentration of fumonisin  $B_1$  in corn of UK (0.2) to 6 mg/kg).

#### **3.4 Africa**

Albeit the majority of African territory has a weather distinguished by high temperature and humidity which is suitable for development of molds, little data is accessible on the occurrence of toxins of *Fusarium*. To a large extent, infection of the primary material is an expanding problem in Africa. Regulative problematic matters are not feasible in the territory of food retailing and exhibition, and mycotoxin issues now have been combined with some food infection in soe parts of Africa [98].

The maximum level of fumonisin B1 in researches on corn of South Africa is very high and achieved to 117.5 mg/kg in Rheeder et al., [67] and 8.514 in new literature by Mngqawa et al. [70].

Getachew et al., [61] detected the fumonisin  $B_1$ ,  $B_2$ , and  $B_3$  in corn of Ethiopia.

Evaluation of Fumonisins on corn products of Africa is low, and these investigations consisted of corn meal [86], corn kernel [94], and corn grits [86].

#### **4. METABOLISM AND MECHANISM OF FUMONISINS**

Structure of fumonisin B has a noticeable similarity to sphinganine and sphingosine. In Fig. 2 both sphingosine and sphinganine are<br>intermediates in the degradation and intermediates in the degradation and biosynthesis of sphingolipids. Furthermore, D'mello et al. [99] reported that fumonisin B obstruct sphingolipid biosynthesis by specifically<br>
inhibiting sphingosine (sphinganine) Ninhibiting sphingosine (sphinganine) Nacyltransferase, *in vitro* and *situ*.

Sphingolipids are a group of lipids which could be detected in the whole of eukaryotic cells. All of the sphingolipids include a sphingoid (long-chain base backbone). Sphingolipids are critical basic molecules and rule as regulators of a numeral of cell act [101]. In Fig. 3 presents location of working of fumonisin B-induced inhibition of the enzyme CER synthase, is presented.

## **4.1 Mechanism of Fumonisins in Apoptosis and Cancer**

Interruption of sphingolipid metabolism can cause the increase in available sphingoid backbone and their 1-phosphates may change the compound sphingolipids and decrease the biosynthesis of ceramide (CER). Available sphingoid backbone induced cell death, but Fumonisins by inhibition of CER synthase can restrain cell death [102].

Feedback of the apoptosis and carcinogenicity effects induced by Fumonisin  $B_1$  can be some mechanisms including oxidative damage, lipid peroxidation and maybe an induction of hepatic and renal tumors happens [16]. Also, Yin et al. [103] discovered that  $FB<sub>1</sub>$  was able to promote the production of free radicals (by increasing the rate of oxidation) and by lipid peroxidation in membranes can accelerate chain reactions.



**Fig. 2. Structures of fumonisin B, sphingosine, sphinganine and ceramide backbone [3,100]**



**Fig. 3. A summarized scheme of the sites of action of fumonisin B-induced inhibition of the enzyme ceramide synthase on the pathway of de novo sphingolipid synthesis and turnover in mammalian cells and [16]**

In some studies following Fumonisin  $B_1$  treatment in different cells of human and animals, it has been proven that apoptosis caused by Fumonisin  $B_1$  does not entail p53 or Bcl-2 group proteins and protect cells from the apoptosis by baculovirus gene (CpIAP). Baculovirus gene obstructs induced apoptosis by the tumor necrosis factor (TNF) pathway that cleaved caspase-8. Probably, the mitochondrial pathway consists of induced apoptosis by Fumonisin  $B_1$  by the actuation of Bid and release cytochrome c [16].

Wang et al. [104] reported that Fumonisin  $B_1$  in human normal esophageal epithelial cells

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(HEECs) stimulated the proliferation, whose mechanism of HEECs is, decreasing in protein expression of cyclin E, p21, and p27 and increase in protein expression of cyclin D1.

### **4.2 Mechanism of Fumonisins in Hepatotoxicity**

Accumulation of sphingoid base because of induced fumonisin  $B_1$  can provoke TNF- $\alpha$  and make the hepatotoxicity in mice. Also, TNF-α receptor 1b is important mediating in the hepatotoxic responses by a rise in the circulation of liver enzymes [105].



**Fig. 4. A schematic landscape of the pathways conduct to apoptosis and the mechanisms probably consisted of fumonisin B1 -induced activation of caspase-3 resulted in apoptosis. X mark showed the mechanisms that do not consisted of the apoptosis caused by fumonisin B1 [16]**

#### **4.3 Mechanism of Fumonisins in Immunotoxicity**

Exposure to  $FB_1$  in human dendritic cells; increases the exhibition of IFN-γ and the associated chemokine CXCL9. Nevertheless, fumonisin  $B_1$  may decline the lipopolysaccharideinduced liver and brain expression of IL-1β and IFN-γ in addition to the induced lipopolysaccharide expression of IL-1β, IL-6, and the chemokines CCL3 and CCL5 in human dendritic cells [16].

In piglets, fumonisin  $B_1$  exposure can increase expression of IL-18, IL-8, and IFN-γ mRNA. But mRNA measure of TNF-α, IL-1β in piglet alveolar macrophages and levels of IL-4 may decrease [106,107].

After exposure to fumonisin  $B_1$  in mouse, a raise expression of TNF-α and interleukin-1β (IL-1β) has been observed in kidney and the liver. Also, FB<sub>1</sub> can raise expression of IFN-y, IL-1α, IL-18, IL-12, IL-10, and IL-6 in the liver of mouse [16].

#### **4.4 Mechanism of Fumonisins in Some Disorder**

Smith et al. [108] recommended that the induced<br>Fumonisin B1 by the destruction of Fumonisin B1 by the destruction of cardiovascular action can be one of the primary elements that trigger the occurrence of equine leukoencephalomalacia through the increase in serum and sphingosine concentrations and myocardial sphinganine.

Interruption of sphingolipid metabolism resulted in  $FB<sub>1</sub>$ , before the pregnancy and during the first trimester may affect folate uptake and cause development of the risk of NTD [109 and 110].

 $FB<sub>1</sub>$  increases sphingosine and/or sphinganine concentrations, reduces the mechanical potency of the left ventricle and blocks L-type Ca channels. Generally, pulmonary edema could be caused by acute left-sided heart failure [111,112].

# **5. TOXICITY OF FUMONISINS**

In the human and various animals, Fumonisins beget some toxic effects such as carcinogenic, hepatotoxic. and nephrotoxic. Moreover, sensitivity to Fumonisins is different in human and varies in animals. For example, based on Bondy et al. [113] rats are more sensitive to fumonisin  $B_1$  than mice. We summarized disorder effects, dosage, duration and source of fumonisin in Table 2.

## **5.1 Carcinogenicity**

Contamination of wheat, corn and rice with Fumonisin B can increase the risk of esophageal cancer in human [16,45,114], by stimulating the proliferation of human esophageal epithelial cells (HEECs) [104]. Furthermore, Mathur et al. [117] observed some different effects of stimulation of the proliferation in liver cells consisted of a proliferation of ductular bile cells and hepatocyte proliferation in cattle.

In rats, continuing intake of  $FB<sub>1</sub>$  (up to 2 years) has consequences such as the introduction of renal tubule tumors, hepatocellular adenomas, cholangiocarcinomas, and carcinomas [130,132].

## **5.2 Hepatotoxic Effect**

Fumonisins could create a mild hepatopathy in lambs [11]. Nonetheless hepatotoxic effects of Fumonisins in cattle is more extensive than lamb, and consists of increases in gamma-glutamyl transpeptidase (GGT), lactate dehydrogenase (LDH), serum aspartate aminotransferase (AST), cholesterol and bilirubin, and mild microscopic liver lesions [116]. Hepatic lesions were distinguished by the different severity of disorganized hepatic cords and hepatocyte apoptosis [117]. Therefore, Therefore, it is possible that cattle is more sensitive to Fumonisins than lamb.

Increasing dietary Fumonisin  $B_1$  increased liver weight, serum calcium, cholesterol, and AST levels. Also, biliary hyperplasia and multifocal hepatic necrosis were present in broiler chicken [8]. In researches of Kubena et al. [121] and Weibking et al. [7] chickens fed with Fumonisin B1, sphinganine: sphingosine (Sa:So) ratio, serum glutamate oxaloacetate aminotransaminase (SGOT), levels of free sphinganine in the serum, AST ratios, LDH, and GGT increased. Nonetheless, total liver lipids of chicks decreased significantly. Subacute treatment of broiler chicks with Fumonisin  $B_1$ <br>bring about hepatic oxidative stress oxidative simultaneously by SA/SO gathering. Also, TBARS (Thiobarbituric acid reactive substance) levels, catalase activity, and Vit C content increased [122]. Therefore, (according to the measure of treatment with Fumonisins) sensitivity to Fumonisin in broiler chicken increased in comparison with the latest research. Additionally, hepatotoxic effects of Fumonisins besides of change in the level of liver enzymes can influence other factors like Vit C content, TBARS, and even liver weight of broiler chicken.



# **Table 2.** *In vivo* **disorder effects induced by Fumonisins**









Feeding the turkey with Fumonisin  $B_1$  caused an increase in liver weight and serum AST levels. However, serum cholesterol, alkaline phosphatase, MCH (mean cell hemoglobin) and MCV (mean cell volume) declined. Likewise, hypertrophy of Kupffer's cells and biliary hyperplasia were present in these turkeys [13].

In ducks, FB1 increased the level of cholesterol, total protein, alanine aminotransferase (ALT), LDH, GGT and SA/SO (sphinganine to sphingosine ratio) in the plasma. Also, FB1 resulted in the growth of liver weight by liver hyperplasia [126]. These effect of Fumonisins in ducks probably created by SA to SO ratio and oxidative damages.

FB1 in mice decreased Ferric reducing antioxidant power (FRAP) content in liver and increased non-protein thiols (NPSH) levels [131] and liver enzymes like AST and ALT in circulation [105]. Moreover, FB1 caused an increase in serum levels of the total bile acids, alkaline phosphatase, and cholesterol, and created microscopic effect such as hepatocellular hypertrophy, hepatocellular apoptosis, Kupffer cell hyperplasia, hepatocellular single cell necrosis, mitosis, anisokaryosis, and macrophage pigmentation [113,128].

Effects of  $FB<sub>1</sub>$  on rabbits are a significant increase in liver weight [135], alkaline phosphatase (AP), total protein, AST, ALT, and GGT. Furthermore, degeneration of hepatocytes and apoptosis were the prominent degenerative changes in the liver of rabbits [14,134].

Because of fumonisin  $B_1$ ,  $B_2$ , and  $B_3$ , hepatic necrosis in ponies occurred [138].

Effects of Fumonisins in the liver of piglet were apoptosis, necrosis, hepatocyte proliferation, hyperplastic hepatic nodules (in chronic studies), icterus, and hepatocellular necrosis. Besides, the serum cholesterol, alkaline phosphatase, AST activities, sphinganine and sphingosine concentrations in kidney, heart, lung, and liver were elevated. However, there were no detectable portal triads or central veins, adjacent parenchyma, and the perilobular connective tissue was compressed [10,142-144]. The hepatic changes especially disorganization in piglet by Fumonisins is probably because of an acute pathway of this mycotoxin.

#### **5.3 Kidney Toxicity**

Fumonisin in the kidney of lambs revealed with tubular nephrosis [11].

Accumulation of sphingosine and sphinganine in the kidney of calves created renal lesion like vacuolar change, karyomegaly, apoptosis, dilatation of proximal renal tubules (that included protein and cellular debris) and the proliferation of proximal renal tubular cells [117].

Effect of fumonisin in the kidney of turkeys and broiler chicken was increasing in kidney weight [7,13,119].

In both sexes of rats, Fumonisins caused decrease in the weight of the kidney, nephrosis in outer medulla (especially in female rats) [12]. Ferric reducing antioxidant power (FRAP) content in the kidney of mice was decreased [131].

Bucci et al. [134] and Orsi et al., [14] reported that the effect of Fumonisin in the kidney of the rabbit was apoptosis and degeneration of renal tubule epithelium, it caused an increase in the level of urea and creatinine, too.

Fumonisins in the kidney of pigs created a mild degenerative change, and in the urine of pigs, the highest Sa/So ratio and Sa ratio were produced in the  $48<sup>th</sup>$  h [9,143].

According to these studies, toxic effects of Fumonisins in the kidney is not extensive such as liver and sensitivity of kidney of rodents and chicken to Fumonisins is lesser than other animals.

#### **5.4 Leukoencephalomalacia**

Fumonisins (especially fumonisin  $B_1$ ) are the causal factor in the development of LEM in horses [137]. The lethality rates, mortality, and morbidity in horses were 85.7%, 10%, and 11.6% respectively [6].

Because of Fumonisins in horses with LEM, brain lesions as the following were observed: severe to early bilaterally symmetrical edema of the brain; brown-yellow discoloration; focal necrosis in the medulla oblongata; focal or multifocal areas of hemorrhage; sporadically pyknotic nucleus all over the parts of rarefaction hemorrhage; softening of the sub-cortical white matter; cavitations crowded with proteinaceous edema with rarefaction of the white matter; mild percolation by infrequent eosinophils and neutrophils; intracytoplasmic eosinophilic globules; inflamed glial cells with plentiful eosinophilic cytoplasm; separation of cell edges;

hyperchromatic; edema; necrosis; large parts of malacia in the white matter of the cerebral hemispheres; cerebellum; brainstem [6,136,137]. These brain lesions (emerged by Fumonisin in horses) is probable to lead horses to show nervous signs. These signs mainly include apathy; incoordination; walking into objects; changes in temperament; paralysis of the tongue and lips in one of the horses; paresis of tongue and the lower lip; inability to drink or eat; a widebased stance; reluctance to move; trembling; hyperexcitability; four leg ataxia; blindness; tetanic convulsion; aimless walking and circling which ends to death [6,136,137].

Fumonisin created leukoencephalomalacia in rabbits, and the bilateral brain microscopic lesions consisted of small focal bleeding in the malacia, cerebral white matter, and bleeding in the hippocampus [134]. However, brain lesions and nervous signs because of leukoencephalomalacia in rabbits, is not as extensive and prevalent as horses. Therefore the brain of horses is more sensitive than rabbits, to Fumonisins.

#### **5.5 Porcine Pulmonary Edema (PPE)**

Usual damage of Fumonisin B in pigs was severe edema in the lung by inhibiting sphingolipid biosynthesis, phagocytosis in pulmonary macrophages, and gathering of substance material in pulmonary capillary endothelial cells [9,144].

The clinical signs in pigs because of pulmonary edema (induced by Fumonisins) consisted of; hydrothorax and respiratory distress (revealed by increasing respiratory rate and effort with open mouth and abdominal breathing). Lethal pulmonary edema appears during 4 to 7 days after the daily feed or intravenous treatment of FB1[10,144].

#### **5.6 Other Toxic Effects**

Exposure to  $FB_1$  during the first trimester and before the pregnancy emerged to developed the hazard of neural tube defects (NTD; because of the defeat of the neural tube to close, embryonic defects of the spinal cord and brain happened) [15,140]. Also, Fumonisins in human colon cells caused to main and early induction of lipid peroxidation, assessing IL-8 secretion, and increasing in membrane microviscosity [115].

Feeding by fumonisin in calves has some effects such as; impairing the lymphocyte blastogenesis [116], decreasing in serum-induced granulosa cell (GC) proliferation [118], lethargy, increasing of sphingosine and sphinganine concentration in the heart, lung, and skeletal muscle. Raise in the concentration of sphinganine, but not sphingosine, in brains of managed calves [117], and have no effects on cell proliferation,<br>progesterone production, CYP11A1 and progesterone production, CYP11A1 and CYP19A1 gene expression of bovine granulosa cell [118].

Diarrhea and lethargy detected in fumonisin administrated lambs [11].

In broiler chicks,  $FB<sub>1</sub>$  had an adverse effect on weight, water consumption, feed efficiency, and body [119]. Also, Fumonisins reduced villus height and crypt depth of the ileum, the abundance of Candidatus Savella and Lactobacilus spp [123], and body weight, but Clostridium perfringens content (reason of subclinical necrotic enteritis), and the weight of bursa of Fabricius, gizzard as well as proventriculus increased. Other effects of  $FB<sub>1</sub>$ were diarrhea, thymic cortical atrophy, rickets [8,119], slightly inhibition in estradiol production [123], and elevation in the level of serum nitric oxide (NO) [124].

Henry and Wyatt. [125] reported that fumonisin in the egg could cause extreme hemorrhages of the thoracic area, head, and neck of the dead embryos in the egg.

Fumonisin  $B_1$  in turkey appeared thymic cortical atrophy, and moderate enlarging of the proliferation and degeneration of hypertrophied zones of tibial physis [13].

In mice, Fumonisins can cause adrenal cortical cell vacuolation and mild to moderate gastric atrophy and may cause an increase in serum cholesterol. Vacuolated lymphocytes and myeloid cells were also detected in mice due to Fumonisins [113]. Also, Fumonisins decreased the number of parietal cells, proliferative activity of gastric glands, gastric mucosa height and mitotic index in the gastric glands [129]. In contrast, Dassi et al. [131] did not detect any change in the indirect nitric oxide (NOx) content, TBARS, ascorbic acid, organ-to-body weight ratio, organ-to-adrenal gland weight ratio or organ-to-brain weight.

Fumonisins in pigs had some effects such as reduction in left ventricular dP/dT (max) (an indicator of heart contractility). But mean pulmonary artery pressure, heart rate, mean systemic arterial pressure, cardiac output, and pulmonary artery wedge pressure (by obstruction

of L-type Ca channels) were increased by get up sphinganine and/or sphingosine mass. Also, in previous studies, parakeratosis, postpone in the pattern of papillary of the distal esophageal mucosa (part of stratum basale), hyperkeratosis, and hyperplastic nodules in the liver cells, esophageal plaques, and right ventricular hypertrophy were detected [141,142,144].

Effects of Fumonisins on hoof cells of horses were increasing in supernatants of explants, and decreasing in lamellar integrity at noncytotoxic concentrations, but Fumonisins didn't influence dermal or epidermal cells [139].

#### **6. DIETARY INTAKE**

In the European diet, the total intake of  $FB<sub>1</sub>$  has been evaluated at 1.4 µg/kg of body weight/week [145]. Daily intake of Fumonisins in different countries and foods, are summarized in Table 3.

In Soriano and Dragacci,. [145] and Creppy., [146] papers, tolerable daily intake (TDI) of  $FB<sub>1</sub>$ was reported 800 ng/kg. Also, provisional-<br>maximum-tolerable-daily-intake (PMTDI) of maximum-tolerable-daily-intake (PMTDI) of fumonisin was noted 2 µg/kg of body weight per day by the no-observed-effect-level (NOEL) of 0.2 mg/kg of body weight/day and a safety aspect of one hundred.

By means of simulation model, mean concentrations of Fumonisin  $B_1$  in milk is evaluated 0.36 μg/kg. However, the pretended TDI from milk for females and males fell below European Union guidelines [147].

Orsi et al. [14] demonstrated that feces are the major way of excretion of fumonisin  $B_1$  in rabbits, by comparing the concentration of  $FB<sub>1</sub>$  in urine, liver, and feces.

#### **7. MAXIMUM LIMITATION**

Different variables may affect the foundation of tolerances for specific mycotoxins, such as delivery of mycotoxins through products; regulation of trade contact in different countries; availability of data of toxicological or dietary exposure; and accessibility of techniques for analysis [148].

Deadline level for Fumonisins in maize and other cereals, at the moment changes from 5 to 100000 µg/kg. Table 4 illustrates present laws of Fumonisins in feeds and foods, set by nations

such as America, Africa, Europe, and Asia and described by [149,150].

#### **8. DETOXIFICATION OF FUMONISINS**

Strategies of detoxification for infected feeds and foods to diminish or remove the toxic effects of Fumonisins by biological, physical, and chemical processes are essential to boost food safety, hinder financial damages, and recover infected commodities. Table 5 includes data detected on biological, physical, and chemical processes for detoxification of Fumonisins in an abbreviated form.

#### **8.1 Biological Methods**

An enzymatic detoxification process of recombinant enzymes from the bacterium *Sphingopyxis sp.* resulted in hydrolysis of Fumonisin  $B_1$  to HFB<sub>1</sub>; deamination of HFB<sub>1</sub> by aminotransferase (a miss of the two tricarballylic side-chains via carboxylesterase) in the existence of pyridoxal phosphate and pyruvate. Lactic acid bacteria such as *Micrococcus luteus* and *Bacillus subtilis* bind to Fumonisin  $B_1$  and Fumonisin  $B_2$ . Peptidoglycan binds to at least one tricarballylic acid part in the structure of  $FB<sub>1</sub>$ and especially  $FB<sub>2</sub>$  [2].

*L. plantarum* MYS6 is having potential probiotic attributes and antifungal activity against Fumonisin producing *F. proliferatum* MYS9 [183].

52.9% of  $FB_1$  and 85.2% of  $FB_2$  were removed by two *Lactobacillus* strains (*L. pentosus X8* and *L. plantarum B7*), in the aqueous medium [165].

Fermentation using three different yeast strains (*Saccharomyces*) is a method for detoxification of Fumonisins. Thus a maximal decrease was observed in 28% and 17% for Fumonisin  $B_1$  and Fumonisin  $B_2$ , respectively [164].

Hydrolyzing ester bonds of fumonisin  $B_1$  by black yeasts (*Exophiala spinifera* and *Rhinoclodiella atrovirensa*) reported by [166].

*Candida parapsilosis* could inhibit mycelial growth of *Fusarium* species from 74.54% and 56.36%, and the maximum and minimum decrease in total created fumonisin was 78% and 12%, respectively [167]. Therefore, we can remove 17 to 85 % of Fumonisins through the biological process, and *Lactobacillus* known as the most effective strains for detoxification of Fumonisins.





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Food	<b>Nation</b>	Intake (ng/kg of bw/day)	<b>Explantion</b>	<b>Reference</b>
Food with corn based	Denmark	400		[81]
Food with corn based	South Africa	14,000 to 440,000	A group of people exhibiting a high prevalence of human esophageal [155]	
Food with corn based	South Africa	5,000 to 59,000	A group of people exhibiting a less prevalence of human esophageal	$[155]$
Food with corn based	UK	30		$[160]$
Corn inferred commodities	Belgium	16.7		$[145]$
Corn inferred commodities	China	450 to 15,810 (Mean=3020)	Camputed on the base of the 50 kg body weight	$[33]$
Corn inferred commodities	Germany	10.4	Users >14 years	[145]
Corn inferred commodities	Italy	185.6	Italian users	[145]
Corn inferred commodities	Italy	24.6	All people in Italy	$[145]$
Corn inferred commodities	Norway	0.24	Adult male and female population	[145]
Corn inferred commodities	Norway	0.50	Adult male and female users	[145]
Corn powder	Argentina	79 to 198	For samples during 1996/1997 and January 1998	[87]
Corn pieces	Germany	69.8	Users >14 years	$[145]$
Corn pieces	Italy	283.6	Italian users	[145]
Corn pieces	Italy	15.9	All people in Italy	$[145]$
Food	Mexico	$0.4(0-23.2)$	User in state of Morelos	[161]
Food	<b>Burkina Faso</b>	$0.8(0-2.4)$	All users	[161]
Food	South Africa	3.8	User in Transkei	[161]
Food	South Africa	0	User in KwaZulu-Natal	[161]
Food	Guatemala	3.5	Urban area	[161]
Food	Guatemala	15.6	Rural area	[161]
Food	Guatemala	$0.2 - 23$	All users	[162]
Rice	France	12.1	All children in france	$[145]$
Rice	France	5.6	All female adults in france	[145]

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Food	<b>Nation</b>	Intake (ng/kg of bw/day)	<b>Explantion</b>	Reference
<b>Rice</b>	France	5.6	All male adults in france	[145]
Rice	France	5.7	All people in france	$[145]$
Rice	Germany	0.6	Users >14 years	[145]
Wheat commodities	France	345.1	All children in france	[145]
Wheat commodities	France	230.8	All female adults in france	[145]
Wheat commodities	France	256	All male adults in france	$[145]$
Wheat commodities	France	240.08	All people in france	[145]
Wheat commodities	Italy	62.1	Italian users	[145]
Wheat commodities	Italy	10.6	All people in Italy	[145]
Food and feeds	Germany	bad case scenario: 21,000 German users		[163]
		mean case scenario:		
		1,100		



# **Table 4. Maximum limits for Fumonisins in feeds and foods in different countries [149,150]**



# **Table 5. Biological , physical , and chemical processes of Fumonisins**



#### **8.2 Physical and Chemical Methods**

Fumonisin  $B_1$  needs a massive temperature (150–200°C) to gain 87–100% demolition in corn cultivation [166].

During extrusion of dry-milled products, reduction of the measure of Fumonisins was 20–50% for non-mixing extruders and 30–90% for mixingtype ones [184]. For the production of cornflakes through the extrusion and roasting of raw corn, 60–70% of Fumonisins  $B_1$  and  $B_2$  were loosened; however, removing of Fumonisins only in the extrusion step was less than 30% [168]. Destruction of Fumonisin  $B_1$  in extrusion process of grits, was 92% [168]. The economic, lowest toxic and most biodegradable solvent for Fumonisin extraction is ethanol-water [173].

In baking corn muffins, removing Fumonisin during the baking process for 20 minutes were amidst 16 and 28% at 175°C and 200°C, respectively. Additionally, flotation of the corn in water reduced the amount of Fumonisin  $B_1$ , and frying corn chips for 15 minutes at 190°C brings about a 67% remove of the Fumonisin. However, spiked corn masa fried at 140–170°C (while degradation begins to take place above 180°C) has no significant loss of Fumonisin  $B_1$  [171 and 185].

One of the most impressive management to cause declination of the measure of Fumonisin B1 is a 0.2% solution of  $SO<sub>2</sub>$  at 60°C for six hours [179]. Nevertheless, canning and cooking had a small influence on fumonisin measure [172].

In paper of Solfrizzo et al. [174] the adsorption capacity of cholestyramine for fumonisin  $B_1$  was 85% from a solution including 200  $\mu$ g/ml FB<sub>1</sub>.

Detoxification of corn with ammonia process reduced fumonisin levels (30 to 45%), and no mutagenic potentials were found in the managed corn [175].

Obstruction in the amine group of fumonisin B1 by reaction with fructose is another way to the detoxification of fumonisin  $B_1$  [176].

The percentage of reduction in  $FB<sub>1</sub>$  in corn by single  $Ca(OH)_2$  (nixtamalization) or with Na- $HCO<sub>3</sub> + H<sub>2</sub>O<sub>2</sub>$  (modified nixtamalization), was 100% [178].

Chlorophorin gets from vanillic acid, ferulic acid, caffeic acid, and iroko decreased  $FB<sub>1</sub>$  levels by 90–91% [177].

Although treatment with oxidizing agents is an economical method for detoxification of Fumonisin  $B_1$ , this method isn't demonstrated in bioassays [178].

The acidic aqueous solution such as  $NaNO<sub>2</sub>$  can create deamination in fumonisin  $B_1$ , significantly [180].

In the floating section after treatment with NaCl solution, 86% of  $FB<sub>1</sub>$  were removed [181].

Celite and O3 couldn't make any significant difference in the level of  $FB<sub>1</sub>$ , but bentonite adsorbed only 12% of the  $FB<sub>1</sub>$  [174,182].

According to these reports, physical and chemical methods are the most effective ways of detoxification of Fumonisin (in comparison with the biological method), so that an intervention is necessary for removing the Fumonisin from feeds and foods.

#### **9. CONCLUSION**

Fumonisins can cause fatal diseases in animals and are classified as a potential human carcinogen. In this paper, we reviewed the aspects of studies concerning the ability of Fumonisins to cause various toxicity effects in different parts of body in human and animal. We evaluated and compared occurrence of Fumonisins in several countries. We also evaluate the effects of different detoxification method for removing the Fumonisins, mechanism of toxicity in cells of human and animals, the intake of Fumonisins in various consumers, and to compare the limitation of Fumonisins in countries mentioned above.

For future investigation about Fumonisins, the authors suggest estimating the reproductive effects of Fumonisins; improving the information about the occurrence of Fumonisins in different parts of the world; extending masked Fumonisins in detoxification researches; improving the legislation about Fumonisins to change daily intake of these mycotoxins; growing attention to mechanisms of Fumonisins in different types of animals and cells; cell-cell interactions; exposure pathways; and finally the exposure measures.

#### **CONSENT AND ETHICAL APPROVAL**

It is not applicable.

#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

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