



Phytochemical Screening and Free Radical Scavenging Activity of Hydroethanolic Leaf Extract of *Senna sieberiana* DC (Caesalpiniaceae) and its Fractions

Alioune Dior Fall^{1*}, Awa Ndiaye Sy², Serigne Ibra Mbacké Dieng¹,
Abdou Sarr¹ and Mbaye Dieng¹

¹Laboratory of Pharmacognosy and Botany, Faculty of Medicine, Pharmacy and Odontology,
Cheikh Anta Diop University, Dakar, Senegal.

²Laboratory of Pharmacology and Pharmacodynamics, Faculty of Medicine, Pharmacy and Odontology,
Cheikh Anta Diop University, Dakar, Senegal.

Authors' contributions

This work was carried out in collaboration among all authors. Author ADF planned all experiments. Authors ADF, SIMD, AS and MD supported the extraction, fractionation and phytochemical screening. Authors ADF, ANS and SIMD carried out the free radical scavenging study and provided the statistical analyses of data. Author ADF wrote the first draft of the manuscript. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/EJMP/2019/v29i330154

Editor(s):

(1) Dr. Patrizia Diana Professor, Department of Molecular and Biomolecular Sciences and Technologies, University of Palermo, Palermo, Italy.

(2) Dr. Marcello Iriti Professor, Plant Biology and Pathology, Department of Agricultural and Environmental Sciences, Milan State University, Italy.

Reviewers:

(1) Evgeny Puchkov, Russian Academy of Sciences, Russia.

(2) Marjan Vracco, national Institute of Chemistry, Slovenia.

(3) Raju Senthil Kumar, Swamy Vivekanandha College Of Pharmacy, India.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/50631>

Received 17 June 2019

Accepted 19 August 2019

Published 11 October 2019

Original Research Article

ABSTRACT

Objectives: The aim of this study was to identify the phytochemical groups of hydroethanolic leaf extract of *Senna sieberiana* DC and its fractions and to investigate their free radical scavenging activity.

Methods: *S. sieberiana* leaves were extracted with hydroethanolic solvent. From the hydroethanolic extract 3 fractions were obtained after a liquid/liquid fractionation

*Corresponding author: E-mail: alioune.fall@ucad.edu.sn; elieufall@yahoo.fr;

(dichloromethane, ethyl acetate and water). Phytochemical screening of the leaf extract and its fractions was done using standard reactions. Free radical scavenging activity was assessed using DPPH and ABTS assays.

Results: Tannins, flavonoids, anthracenic derivatives, sterols and triterpenoids were the main phytochemical constituents of the leaf extract and fractions. The hydro-ethanolic leaf extract of *Senna sieberiana*, its dichloromethane, ethyl acetate and aqueous fractions and ascorbic acid had respective IC₅₀ values of 191.6±3.82 - 495.73±8.96 - 165.8±4.85 - 50.40±2.65 - 19.53±0.13 µg/ml in ABTS assay. In DPPH assay, the IC₅₀ values were 44.8 ± 1.22 - 218.93±9.01 - 32.13 ± 1.8 - 26.4±0.11- 4.66±0.07 µg/ml respectively for the leaf extract, dichloromethane, ethyl acetate and aqueous fractions and ascorbic acid.

Conclusion: The hydroethanolic leaf extract of *S. sieberiana* had shown free radical scavenging activity. The aqueous fraction was more active among plant tested samples.

Keywords: *Senna sieberiana*; leaf extract; fractions; phytochemical; scavenging activity.

1. INTRODUCTION

Senna sieberiana DC (synonym: *Cassia sieberiana* DC) is a plant widely spread in West Africa. Its leaves are used in Senegalese traditional medicine as detoxing, antipyretic, diuretic and against anemia [1]. Leaf extract are also used to treat various diseases such as stomach ache, ulcer and diarrhoea.

Leaf extract of *Senna sieberiana* have been reported for their antimicrobial, antitrypanosomal, antiviral and anthelmintic properties [2-5].

Oxidative stress due to overproduction of free radicals has been involved in neurodegenerative disease (Alzheimer and Parkinson diseases), diabetes, cardiovascular disease, atherosclerosis, rheumatoid arthritis [6,7]. What makes us to assess the free radical scavenging activity of hydroethanolic leaf extract of *S. sieberiana* and its fractions.

2. MATERIALS AND METHODS

2.1 Plant Collection

Leaves of *Senna sieberiana* DC were collected at the Botanical Garden of the Faculty of Medicine, Pharmacy and Odontology (Cheikh Anta DIOP University, Dakar / Senegal). The plant was identified and authenticated by Dr W. Diatta (Herbarium of the Botanical Garden of the Faculty of Medicine, Pharmacy and Odontology of Dakar) where a voucher specimen was kept. Plant leaves were air dried at room temperature. Dried leaves were ground to a fine greenish powder.

2.2 Extraction and Fractionation

An amount of 125 g of powdered leaves of *S. sieberiana* was decocted twice for 30 minutes using one liter of ethanol/water (80:20, v/v) and filtered through Whatman No. 1 filter paper. The solvent was removed under reduced pressure using a rotary evaporator in order to get the hydroethanolic leaf extract.

For liquid/liquid fractionation, 2.5 g of dried leaf extract was dissolved in a mixture (distilled water/ dichloromethane; 1:1). After decantation in a separatory funnel, the aqueous solution obtained was extracted again twice with dichloromethane. The dichloromethane solutions were combined and evaporated to give the dichloromethane fraction. The aqueous solution was again subjected to liquid-liquid extraction with ethyl acetate under the same conditions as above. The ethyl acetate and aqueous solutions obtained were evaporated separately and lead to the corresponding fractions.

2.3 Phytochemical Screening

To test for the presence of phytoconstituents groups, standard phytochemical analyses were carried out on the hydroethanolic leaf extract and its fractions. Chemical tests were carried out on these samples using standard procedures for the detection of condensed and hydrolysable tannins (Stiasny test followed by ferric chloride test), flavonoids (Shibata's test), anthracenic derivatives (Borntraeger test), cardiac glycosides (Baljet, Kedde and Raymond-Marthoud reagents tests), steroids and triterpenoids (Liebermann-Buchard test), carotenoids (antimony chloride/chloroform test), alkaloids (Bouchardat, Valser-Mayer and Dragendorff's reagents tests),

in order to identify the presence of phytochemical constituents [8].

2.4 Free Radical Scavenging Activity

2.4.1 ABTS assay

Reduction of free radical ABTS (2, 2-azinobis-3ethylbenzothiazoline-6-sulfonic acid) was investigated using the described method [9]. Two stock solutions of 7.4 mM ABTS and 2.6 mM potassium persulfate were prepared and mixed in equal volumes before allowing them to react for 12 h at room temperature in darkness. This mixture was diluted by adding ethanol, in order to obtain an absorbance of 0.7 at 734 nm. Samples (2 ml) were mixed with 2 ml of ABTS solution and the mixture was left at room temperature for 2 h in darkness. The absorbance of each sample was measured at 734 nm after 30 min by spectrophotometric method.

Experiments were done in triplicate and the ABTS free radical scavenging effect was expressed as IC₅₀ (concentration of sample required to scavenge 50% of free radicals).

2.4.2 DPPH assay

The determination of the DPPH free radical scavenging activity of samples was done using the described method [10]. An ethanol solution of DPPH was prepared by dissolving 4 mg in 100 ml of ethanol. An aliquot of each sample (0.8 ml) at appropriate concentration was added to 3 ml of ethanol solution of DPPH.

The hydroethanolic leaf extract of *S. sieberiana*, its fractions (dichloromethane, ethyl acetate and water) and ascorbic acid were tested at different concentrations. The absorbance of each sample was measured at 517 nm after 30 min. Each experiment was done in triplicate and the absorbance of the initial ethanol DPPH solution did not change after 30 min. The DPPH free radical scavenging effect was expressed as IC₅₀.

2.4.3 Statistical analyses

Data were expressed as mean ± SEM. Analyses of variance (ANOVA) were done for the comparison of results using Fischer's test. Statistical significance was set at $p < 0.05$.

3. RESULTS

3.1 Extraction and Fractionation

From 125 g of dried powdered leaves, 21.87 g of dried leaf extract were obtained corresponding to

a yield of 17.5%. The dichloromethane, ethyl acetate and water fractions represented respectively 12.74 - 31.79 and 54.46% of the dried hydroethanolic leaf extract.

3.2 Phytochemical Screening

Phytochemical screening revealed that anthracenic derivatives, hydrolysable and condensed tannins, flavonoids, sterols and triterpenoids were identified in the hydroethanolic leaf extract of the plant. Negative reactions were obtained for the presence of carotenoids, alkaloids and cardiac glycosides.

The dichloromethane fraction contained anthracenic derivatives, sterols, triterpenoids and condensed tannins. Anthracenic derivatives, condensed and hydrolysable tannins, flavonoids were found in the ethyl acetate fraction while in the aqueous one flavonoids, hydrolyzable and condensed tannins were identified (Table 1).

Table 1. Phytochemical groups identified in leaf extract of *S. sieberiana* and its fractions

Phytochemical groups	HE	DF	EAF	AF
Alkaloids	-	-	-	-
Anthracenic derivatives	+	+	+	-
Cardiac glycosides	-	-	-	-
Carotenoids	-	-	-	-
Flavonoids	+	-	+	+
Hydrolyzable tannins	+	-	+	+
Condensed tannins	+	+	+	+
Sterols and triterpenoids	+	+	-	-

HE: hydro-ethanolic extract, DF: dichloromethane fraction, EAF: ethyl acetate fraction, AF: aqueous fraction

3.3 Free Radical Scavenging Activity

3.3.1 ABTS assay

The hydroethanolic leaf extract of *Senna sieberiana* had an IC₅₀ value (191.6±3.82 µg/ml) higher than those of ethyl acetate and aqueous fractions (respective IC₅₀: 165.8±4.85 µg/ml and 50.40±2.65 µg/ml/ml) ($p < 0.05$). The dichloromethane fraction had exhibited the highest IC₅₀ value (495.73±8.96 µg/ml). Ascorbic

acid had shown the lowest IC₅₀ value (19.53±0.13 µg/ml) (*p*<0.05) (Fig. 1).

3.3.2 DPPH assay

The aqueous fraction had shown the lowest IC₅₀ value (26.4±0.11 µg/ml) among plant samples, followed by the ethyl acetate fraction (IC₅₀: 32.13 ± 1.8 µg/ml) and the hydroethanolic leaf extract (IC₅₀: 44.8 ± 1.22 µg/ml) (*p*<0.05). Dichloromethane fraction (IC₅₀: 218.93±9.01 µg/ml) had revealed the highest IC₅₀ value among plant samples. For ascorbic acid, an IC₅₀ value of 4.66±0.07 µg/ml was obtained (*p*<0.05) (Fig. 1).

4. DISCUSSION

Leaves of *S. sieberiana* contained mainly phenolic compounds such as flavonoids, tannins and anthracenic derivatives [11]. The polyphenol extraction is a crucial step for the valorization of these active principles recognized for their scavenging effect [12]. The appropriate choice of solvents preserving the biological properties of these phenolic constituents is very important [13].

As part of our study, we first carried out a decoction using a mixture of two polar solvents such as ethanol and water. The solvents had ability to extract polar compounds represented in *S. sieberiana* leaves. However ethanol is also able to extract non polar constituents such as sterols, triterpenoids and aglycones represented in the leaves of *S. sieberiana*. The yield of this extraction was 17.5%. From leaves of *Senna alata*, it has been recorded after hydroethanolic maceration an extraction yield of 12.5% [14]. The difference between these values may be due to the process.

Subsequently the liquid-liquid fractionation allowed us to successively obtain three fractions: a dichloromethane fraction containing non-polar compounds; an ethyl acetate fraction with compounds of intermediate polarity, and an aqueous fraction containing the most polar compounds. The lowest yield relatively to the hydro-ethanolic extract was that of the dichloromethane fraction followed by those of the ethyl acetate and aqueous fractions.

These results suggest that the bioactive constituents most represented in the leaves of *S. sieberiana* were mainly polar compounds.

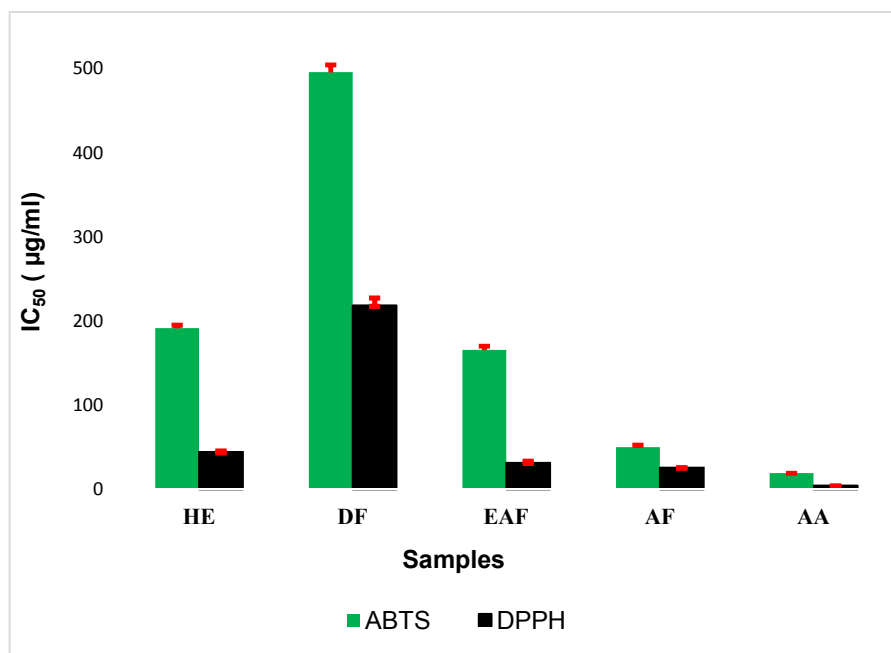


Fig. 1. IC₅₀ (µg/ml) values of different samples in DPPH and ABTS assays
 HE: hydroethanolic extract, DF: dichloromethane fraction, AF: ethyl acetate fraction, AF: aqueous fraction, AA: ascorbic acid

The aqueous and ethyl acetate fractions had shown better capacity to scavenge free radicals, both in ABTS and DPPH assays, than the hydroethanolic leaf extract and its dichloromethane fraction. Among plant samples, the aqueous fraction was seen to be the more active while the dichloromethane fraction exhibited the lowest ability to scavenge the free radicals. What makes us to suggest that polar phytoconstituents, identified in ethyl acetate and aqueous fractions had better ability to scavenge the free radicals. The phytochemical screening had revealed the presence of flavonoids and hydrolysable tannins in ethyl acetate and aqueous fractions and not in the non-polar fraction (dichloromethane fraction). These phenolic compounds such as flavonoids and tannins contained in these fractions could be responsible for the free radical scavenging activity. Indeed polyphenolic compounds are known for their scavenging ability [15,16].

It has been established that the antioxidant efficiency of a proton-bound A radical (AH) increases if the binding force A-H is low and the resulting radical A is as stable as possible. This is the case for phenolic compounds such as flavonoids which are among the best electron or proton donors [17].

Besides in the dichloromethane fraction had been detected condensed tannins which were also found in the ethyl acetate and aqueous fractions. The low ability of the dichloromethane solvent to extract polar phytoconstituents such as polyphenols would explain its low free radical scavenging activity in DPPH and ABTS assays.

5. CONCLUSION

The hydroethanolic leaf extract of *S. sieberiana* which contained tannins, flavonoids, anthracenic derivatives, sterols and terpenoids had exhibited free radical scavenging activity. The ethyl acetate and water fractions were seen to be more active than the leaf extract and the dichloromethane fraction. Aqueous fraction had shown better ability to scavenge free radicals among plant tested samples.

CONSENT

As per international standard or university standard, patient's written consent has been collected and preserved by the author(s).

ETHICAL APPROVAL

As per international standard or university standard, written approval of Ethics committee

has been collected and preserved by the author(s).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Pousset JL. Plantes médicinales Africaines, Utilisation pratiques. Ed. ACCT, Ellipses, Paris. 1989:156.
2. Asase A, Kokubun T, Grayer RJ, Kite G, Simmonds MSJ, Oteng-Yeboah AA, Odamtten GT. Chemical constituents and antimicrobial activity of medicinal plants from Ghana: *Cassia sieberiana*, *Haematostaphis barteri*, *Mitragyna inermis* and *Pseudoecdrella kotschyi*. *Phytotherapy Research*. 2008;22(8):1013-1016.
3. Hoet S, Opperdoes S, Brun R, Adjakidjé V, Quetin-Leclercq J. *In vitro* antiparasitodal activity of ethnopharmacologically selected Beninese plants. *Journal of Ethnopharmacology*. 2004;91(1):37-42.
4. Silva O, Barbosa S, Diniz A, Valdeira ML, Gomes E. Plant extracts antiviral activity against Herpes simplex virus 1 and African swine fever virus. *International Journal of Pharmacognosy*. 35(1);12-16:2008.
5. Waterman C, Smith RA, Pontiggia L, Der Marderosian A. Anthelmintic screening of Sub-Saharan African plants used in traditional medicine. *Journal of Ethnopharmacology*. 2010;127(3):755-759.
6. Valko M, Leibfritz D, MoncolJ, Cronin MTD, Mazur M, Telser J: Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry and Cell Biology*. 2007;39:44-84.
7. Chen X, Guo C, Kong J. Oxidative stress in neurodegenerative diseases. *Neural Regeneration Research*. 2012;7(5):376-785.
8. Joslyn MA. *Methods in food analysis*. Physical, chemical and instrumental methods of analysis. Academic Press, London and New York; 1970.
9. Dudonne S, Vitrac X, Coutière P, Woillez M, Merillon JM. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of Agriculture and Food Chemistry*. 2009;57(5):1768-74.

10. Molyneux P. The use of the stable free radical diphenylpicryl-hydrazyl (DPPH) for estimating antioxidant activity. Songklanakarin J. Sci. Technol. 2004; 26:211–219.
11. Awomukwu DA, Nyananyo BL, Ikpeama AI, Adieze CU. Pharmacognostic Importance in South Eastern Nigeria. Science Journal of Chemistry. 2015;3(3):40-49.
12. Bonnaillie C, Salacs M, Vassilova E, Saykova I. Etude de l'extraction de composés phénoliques à partir de pellicules d'arachide (*Arachis hypogea* L.) Revue de Génie Industriel. 2012;7:35-45.
13. Babbar N, Oberoi HS, Sandhu SK, Bhargav VK. Influence of different solvents in extraction of phenolic compounds from vegetable residues and their evaluation as natural sources of antioxidants. J Food Sci Technol. 2014;51(10):2568–2575.
14. Pieme CA, Penlap VN, Nkegoum B, Taziebou PCL, Tekwu EM, Etoa FX, Ngongang J. Evaluation of acute and subacute toxicities of aqueous ethanolic extract of leaves of *Senna alata* (L.) Roxb (*Caesalpinaceae*). African Journal of Biotechnology. 2006;5(3):283-289.
15. Álvarez R, Araya H, Navarro-Lisboa R, Lopez de Dicastillo C. Evaluation of polyphenol content and antioxidant capacity of fruits and vegetables using a modified enzymatic extraction. Food Technol Biotechnol. 2016;54(4):462-467.
16. Tresserra-Rimbau A, Lamuela-Raventos RM, Moreno JJ. Polyphenols, food and pharma. Current knowledge and directions for future research. Biochem Pharmacol. 2018;156:186-195.
17. Leopoldini M, Marino T, Russo N, Toscano M. Antioxidant properties of phenolic compounds: H-atom versus electron transfer mechanism. J Phys Chem A. 2004;108 (22): 4916-4922.

© 2019 Fall et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/50631>